Emerging Strategies to Attack Polyploid Cancer Cells

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Abstract

Polyploid cancer cells can arise de novo in tumors or they can be induced by therapeutics that inadvertently increase the rate of cytokinetic failure. These cells portend a poor outcome in many cancers because polyploid cancer cells can undergo error prone reductive cell divisions to yield aneuploid progeny. The immune system has evolved mechanisms by which it can specifically recognize and remove polyploid cancer cells, but these appear to be tampered with malignancy so that polyploid cells can persist and fuel the development of cancer cell clones that are resistant to therapeutics and have metastatic potential. Here we review mechanisms by which polyploid cancer cells can arise, are surveilled by the immune system and therapeutic strategies that might prevent or directly attack polyploid cancer cells.

Keywords: Polyploid, Mitosis, Therapeutics, Apoptosis, Cancer, Immune surveillance

Normal Polyploid Cells

While cells with a 2n complement of chromosomes are defined as diploid, cells that possess greater than 2n are referred to as polyploid. The additional DNA content of polyploid cells may seem highly divergent, but polyploid cells exist in mammals and serve vital roles in both development and tissue homeostasis [1]. Placental syncytiotrophoblasts, for example, form the interface between the maternal blood and embryonic fluid. These cells enable gas and nutrient exchange and produce hormones that maintain pregnancy. They are multinucleate cells and are formed and maintained through fusion of the underlying diploid cytotrophoblast cells [2]. Polyploidization can alternatively occur when the genome is replicated but cells fail to undergo cytokinesis. This situation occurs with the maturation of megakaryocytes (MKs). They develop from diploid, bone marrow hematopoietic stem cells, but during maturation they become polyploid by endoreplication, the replication of DNA without cell division. This process is driven by the hormone Thrombopoietin and can result in up to 64n DNA content [3]. The final maturation step for MKs requires extrusion of nuclear content and the formation of proplatelet structures from their cytoplasm. Although still debated, the requirement for polyploidy in MK development is likely related to the requirement for large quantities of mRNA and protein that is eventually packaged into platelets for clotting and repair.

Syncytiotrophoblasts, formed through cell fusion, and MKs, through endoreplication, represent two distinct, specialized cell types where polyploidy plays a vital physiological role. However, aberrant polyploid cancer cells can also arise when the normal checks and balances on cell division are compromised in pseudodiploid cancer cells. Below, we outline how polyploid cancer cells arise and the rationale for attacking this population of cancer cells.

Polyploid Cancer Cells

Oncogenic changes that occur in cancer facilitate mitotic slippage and cytokinetic failure. This disruption facilitates aneuploidy, a numerical change in a fraction of the diploid set of chromosomes. For example, loss-of-function mutations in tumor suppressors such as BRCA2 [4], TP53 [5] and APC [6], all increase the rate of cytokinetic failure, while activating kinase mutations can impact the fidelity of mitosis. Signaling cascades converge to influence the biogenesis and function of centrosomes, the integrity of...
the mitotic spindle assembly checkpoint (SAC), and the completion of cytokinesis [7-9]. The SAC acts as a safeguard for the accurate segregation of chromosomes, ensuring proper attachment of kinetochores to microtubules of the mitotic spindle and optimal tension between bi-oriented sister kinetochores before the transition to anaphase (for review see [10-12]). Defects in chromosome segregation are thought to result from bypass of the SAC. Thus, prior oncogenic events can serve as a prelude to further intratumoral genetic heterogeneity by promoting aneuploidy. This fuels the emergence of more aggressive cancer cell clones over time. Single cell sequencing technologies continue to reveal enormous depth in the clonal heterogeneity found in cancer [13]. However, a prominent pathway to aneuploidy may involve a polyploid intermediate. Polyploidy differs from aneuploidy. Polyploidy is a numerical change in the whole set of chromosomes, not just a fraction. Cancer cells can undergo a viable mitosis but then fail to complete cytokinesis, resulting in the formation of a multinucleate cell. In contrast with the conventional view of polyploidy as a proliferatively arrested state, accumulated data indicate that polyploid cells can undergo reductive divisions that may be error prone, resulting in highly aneuploid progeny that are viable and proliferative [14]. Compared with diploidy, polyploidy serves as a resilient intermediate for aneuploidy because the increased DNA content buffers the loss of essential chromosomes more effectively [15].

Figure 1: Prophylactic and therapeutic strategies for polyploid cells. Illustrated are steps in the malignant progression of cancer and potential therapeutic strategies against polyploid cancer cells. Polyploid cancer cells are a precursor to aneuploid cancer cells through error-prone reductive cellular division and are subject to immune surveillance. Polyploid cells seed the constantly evolving genomic heterogeneity within tumors, spawning clones that are drug refractory and/or have metastatic potential. Therapeutics mentioned in this review that interfere with this process are shown. 1. Therapeutics that attack mitotic cells can induce apoptosis in some cancer cells but also can provoke a subpopulation of polyploid cancer cells that escape from apoptosis (red). 2. Combining flavopiridol (CDK inhibitor) with paclitaxel induces G1 arrest that minimizes development of polyploidy (blue). 3. Combination therapies can alter the propensity for polyploidy by enhancing mitotic catastrophe-induced death of cycling cancer cells (orange). 4. Treatments can show enhanced propensity to attack newly generated polyploid cancer cells, presumably due to vulnerabilities inherent in polyploid cancer cells (green) 5. Inhibition of the interaction between Bcl2 and Beclin1 can enable lethal autophagy as a means to directly attack polyploid cancer cells (purple). Note: A lower case i denotes an inhibitor, i.e. Bcl2i is an inhibitor of Bcl2, such as a Bcl2 mimic; VCR is vincristine.
Cells that are 4n, 8n or more are present in many tumors and the presence of polyploid cells is recognized as a poor prognostic indicator in multiple cancers [16-18]. For leukemias, in particular, it has long been recognized to portend poor outcome [19]. So, the polyploid pool of tumor cells can serve as a constant source upon which cells with variable genomic alterations can emerge, producing therapeutically resistant cells and cells with enhanced metastatic potential over time [14,15,20] (outlined in Figure 1).

Polyploid cells appear particularly well suited to seed metastatic recurrence. One response to polyploidy is the development of cellular senescence [21]. As quiescent cells, polyploids may be uniquely able to survive in the face of chemotherapeutics that target dividing cells. In addition, DNA damage response genes are rewired in polyploid cells, triggering single-stranded base repair and non-homologous end-joining pathways to increase DNA repair activity [22]. The emergence of polyploid cells from senescence to produce viable aneuploid progeny could contribute to tumor recurrence long after chemotherapy has ceased. Multiple lines of evidence suggest they do, in fact, emerge and produce aggressive cancer cell clones [14,20,23,24]. One not fully understood aspect of polyploid cancer cell biology is the observed enhancement in polyploids of properties that are associated with highly malignant cancers, even when these properties are not evident in the associated diploid cancer cell pool. Examples include altered expression of cell cycle regulators [22,25] and markers associated with epithelial to mesenchymal transition (EMT) and cancer stem cells [26,27]. A detailed review of the enhancement of malignant properties in polyploid cancer cells has been published [28].

Importantly, polyploid cancer cells have the ability to seed tumorigenesis, so they do have cancer stem cell-like properties. Polyploid cells isolated from ovarian cancer cell lines express higher levels of the ovarian cancer stem cell marker CD133, form spheroids in culture and tumors in immunocompromised mice [27]. Perhaps most telling regarding the dedifferentiation phenotype of these cells is that they can be selectively manipulated in cell culture to take on properties of mesenchymal lineages from adipose, cartilage and bone [27]. The propensity to undergo EMT has long been considered a factor that portends metastatic dissemination, so the inheritance of developmental plasticity may be an important characteristic that daughter cells inherit from a polyploid precursor.

Such inheritance might be epigenetic. There is evidence, at least in p53 positive cancer, that epigenetic changes in polyploid cells enable silencing of p53 transcripitional targets that activate apoptosis and cell cycle arrest. For example, the DNA methylation inhibitor 5-aza-2-deoxycytidine (5-AzagC) can restore expression of the p53 target and cyclin-dependent kinase inhibitor, p21(CIP), and also restore polyploid cancer cell sensitivity to TNFα [22]. The ability of epigenetic alterations to be passed on to the aneuploid progeny of polyploids has not been sufficiently explored and may contribute to the emergence of drug resistance and metastasis.

**Immunosurveillance**

In immunocompetent mice, tumors that eventually emerge following polyploid cell engraftment are comprised mainly of pseudodiploid cancer cells, so they arise from the progeny of a reductive cell division [29]. The ability of the immune system to specifically detect polyploid cells could be a mechanism that necessitates this reduction in ploidy.

Mechanisms by which the immune system eliminates polyploid cancer cells arise through stress signaling. The protein calreticulin becomes relocatalized to the plasma membrane of polyploid cancer cells, where it acts as a ligand for LDL-receptor-related protein (LRP) (also known as CD91) on the surface of phagocytic cells [30]. To act as an “eat me” signal, translocation must occur from the endoplasmic reticulum (ER) where calreticulin normally functions as a molecular chaperone [31]. Strong evidence that calreticulin is central to polyploid cell immunosurveillance comes from experiments demonstrating calreticulin exposure on the cell surface of polyploid cells does not limit tumorigenesis in immunodeficient mice, but does limit tumorigenesis in mice with an intact immune system [29,32]. Constitutive ER stress in polyploid cells directs calreticulin to the cell surface as manipulations that alleviate ER stress also reduce calreticulin transport to the cell surface and immunogenicity [29].

Polyploid cells are also subject to heightened immunosurveillance by Natural Killer (NK) cells. Hyperploidy-inducing chemotherapeutics induce cell surface expression of ligands for the NK cell-activating receptors NKG2D and DNAM-1 [33]. Again, the ER stress response plays a role. The NKG2D ligand, MICA is upregulated on the surface of both HCT-116 colon cancer cells and K-562 myelogenous leukemia cells by ER stress and this triggers the cytolytic activity of NKS [33]. Polyploidy may therefore prime anti-tumor immunity. Despite this constant immunosurveillance, cancers are often diagnosed at an advanced stage with aneuploidy, dissemination and innate ability to avoid immune effector cells. Despite the advent of immune checkpoint modulating antibodies, restoring the ability of the immune system to identify and attack cancer cells is still a major clinical challenge.

**Chemotherapy-Induced Polyploidy**

Added complexity stems from the fact that therapies

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that attack dividing cells can inadvertently enhance the development of polyplody. For example, drugs that disrupt the mitotic spindle induce prolonged mitotic arrest leading to mitotic catastrophe-induced apoptosis. However, sporadic cells escape, perhaps by falling beneath the threshold of inductive cues required for apoptosis. These cells can alternatively fail to complete cytokinesis, becoming polyplody. Successive rounds of replication with failed cytokinesis can yield cells with even greater than 4n ploidy and while apoptotic signaling is still viable in tetraploid cancer cells induced by nocodazole [34], tetraploid cell lines are more resistant to radiation and DNA damage-induced death than their diploid counterparts [35]. This suggests polyplodyization goes hand-in-hand with dampened ability to activate intrinsic apoptotic signaling. Distinct from apoptosis, enhanced autophagic flux in polyplody cancer cells has been found to either promote or suppress their long-term survival in a context-dependent manner [36,37]. The persistence of polyplody cancer cells is likely enabled by the convergence of multiple escape mechanisms.

Several classes of cancer therapy induce a polyplody cell population, including the clinically used taxanes such as docetaxel [24,38] and paclitaxel, [39], DNA damaging agents such as doxorubicin [40], radiation [14,41] and oncoprotein-targeting compounds [7,42]. This also includes targeted kinase inhibitory drugs that directly attack the mitotic machinery, such as Aurora kinase inhibitors [37,43] and Polo-like kinase inhibitors [44,45]. If cycles of polyplodyization followed by reductive cell division fuel the progressive generation of aneuploidy, then a therapeutic that targets polyplody cancer cells may be a means to short circuit this cycling, attack the evolvability of the cancer genome and enhance the overall effectiveness of many currently used therapies.

Preventing the Development of Polyplody Cancer Cells

In theory, reducing conversion of diploid cancer cells to polyplody cells could be a strategy to limit evolvability. Several lines of evidence suggest combination therapies that work toward this goal. One combination therapy that effectively targets polyplody cell formation, at least in cultured diffuse large B cell lymphoma (DLBCL) cell lines, is the combined use of the histone deacetylase complex (HDAC) inhibitor, Belinostat, alongside the vinca alkaloid, vincristine [46]. Vincristine alone, like other spindle toxins, has a propensity to induce some polyplody alongside mitotic arrest and apoptosis. Belinostat potentiates the apoptotic response. The authors speculate that there are fewer polyplody cells as fewer cells undergo prolonged arrest, mitotic slippage and cytokinesis failure [46]. More cancer cells succumb to acute apoptosis. So, the cooperative effect of these two drugs attacks cells before they have a chance to undergo endoreduplication.

Flavopiridol, a broad spectrum cyclin-dependent kinase (CDK) inhibitor, has also been suggested to reduce the propensity for polyplody cell formation with spindle toxins [47]. This activity is attributed to G1 arrest of cancer cells and occurs even in cells deficient for tumor suppressor genes that abrogate the G1 checkpoint response and which have a propensity for endoreduplication upon treatment with spindle toxins. So, a cytostatic effect of flavopiridol may inhibit endoreduplication and polyplody elicted by spindle toxins, at least in vitro. The propensity for apoptosis to occur alongside flavopiridol-induced G1 arrest can be dependent on both cell type and drugs that are used alongside this CDK inhibitor (for review see [48]).

Development and maintenance of polyplody may come with exploitable energy costs. Polyplod cells have increased size and DNA content and sustaining this while initiating new rounds of DNA synthesis requires increased energy input compared to diploid counterparts. As a master regulator of cellular energy use, mechanistic target of rapamycin complex 1 (mTORC1) translates metabolic and environmental cues into a cascade of events that enable anabolic processes such as mRNA translation and lipid synthesis and can limit catabolic processes such as autophagy. The anti-cancer effects of Aurora kinase B inhibitors are enhanced by co-treatment with mTOR inhibitors [49]. Both rapamycin and torkinib (PP242) potentiated Aurora kinase inhibitor-induced apoptosis and induction of autophagic death in polyplody acute myeloid leukemia (AML) cells. Glycolytic metabolism was found to be enhanced in polyplody cells and cooperation was attributed to enhanced metabolic stress [49]. Along similar lines, activation of 5' AMP-activated protein kinase (AMPK), a direct upstream inhibitor mTOR, by either the natural product, resveratrol, or by salicylate, the active product of Aspirin, can inhibit polyplody cell formation [50]. This occurred alongside treatment with polyplody-inducing drugs nocodazole, cytochalasin D or an Aurora kinase B inhibitor. Importantly, the anti-polyplody activity was validated in vivo using the APCmin model of colorectal cancer [50].

Attacking Existing Polyplody Cancer Cells

Preferentially targeting polyplody cells or preventing the polyplody to aneuploid cell transition could disable tumor progression. High-throughput screening for compounds that selectively kill polyplody cells suggests gene dosage may be an exploitable trait [51]. For example, 8-azaguanine, a compound that requires conversion to a bioactive metabolite by the enzyme hypoxanthine phosphoribosyl transferase 1 (HPRT1), is more toxic to polyplody cancer cells. The extra copies of HRPT1 in polyplody cells underly this toxicity [51]. Altered expression of other genes may
also be exploitable. Genes that regulate meiotic cell division have been found to be upregulated in polyploid cancer cells, alongside genes that regulate mitotic division [41,52]. This means cell division, including the reductive divisions that produce aneuploid progeny, either through nuclear budding, multipolar division or other means, may exploit a distinct set of cell division proteins compared to diploids. It is unknown if any meiosis-specific proteins are absolutely required or druggable in polyploid cells, but identifying such vulnerabilities will move a step closer to targeted therapies for polyploid cancer cells.

A distinct strategy has been uncovered using a cell culture system to examine the synthetic lethality between MYC and inhibition of Aurora kinase B. Pro-survival members of the Bcl2 family were confirmed to enable the persistence of polyploid cells [53,54]. Cooperation between Aurora kinase B inhibitors, and inhibitors of pro-survival Bcl2 proteins, has been previously explored [55-57]. Cooperative effects have been assumed due to enhanced apoptosis through activation of the intrinsic apoptotic pathway. However, new findings suggest a distinct mechanism. Pro-survival Bcl2 family proteins also interact with the BH3 only protein Beclin1 (also ATG6) at the endoplasmic reticulum to blockade autophagy [58,59]. This interaction was demonstrated to be crucial for preventing the lethal autophagy that accompanies polyploidy and to contribute to drug resistance in an in vitro model [54]. This research pinpoints a targetable mechanism of action to directly attack polyploid cancer cells. BH3 mimetic drugs disrupt the interaction of pro-survival Bcl2 family proteins with the BH3 domain of Beclin1 and this tactic can be used in combination with drugs such as Aurora kinase inhibitors to enhance cell killing. BH3 mimetics have also been shown to be effective alongside other drugs that induce polyploidy [60].

Other means to disrupt the Beclin1/Bcl2 interaction may also prove valuable. Ceramides are a family of lipids composed of sphingosine and a fatty acid chain. They are found in various cellular membrane compartments, including the Golgi and lysosome and can modify cell signaling pathways. Short-chain ceramides have been found to induce the dissociation of the complex formed between Beclin1 and Bcl2 through the activation of c-Jun N-terminal kinase 1 (JNK1) [61]. JNK1 phosphorylates the Bcl2 protein and this interferes with the association between Beclin1 and Bcl2, thereby enabling autophagy [61]. For polyploid cells, this autophagy is lethal. Knockdown of the gene encoding the ceramide transport protein (known as COL4A3BP or CERT), which moves ceramide from the endoplasmic reticulum (ER) to the Golgi apparatus, induces expression of lysosome-associated membrane protein 2 (LAMP2) and increases autophagic flux, leading to polyploid cell death [62]. So, COL4A3BP may be a target for therapeutic intervention to attack polyploid cancer cells that ultimately works via disruption of the Beclin1/Bcl2 interaction.

Direct targeting approaches do not only have to target polyploid cells. For example, inhibition of PLK1 alongside treatment with spindle toxins leads to enhanced apoptosis of both diploid and polyploid cancer cells, but polyploid cells have enhanced sensitivity [63]. The enhanced effect of PLK1 inhibition on cells with >4n DNA content was attributed to an inability of polyploid cells to tolerate any further increase in ploidy that was induced by PLK1 inhibition. Polyploid cells were more readily moved toward mitotic catastrophe-induced apoptosis. Genome duplication also increases sensitivity to pharmacological inhibitors of mitotic kinesin family member 11 (also known as Eg5) [64] and monopolar spindle protein 1 (MPS1) [65], so sustained inhibition of mitotic regulators is more toxic to polyploid cells than their diploid counterparts. In theory, these approaches will target both diploid and polyploid cancer cells and could be effective therapies for attacking all cancer cells.

Summary

Genomic instability is a hallmark of cancer and polyploid cells have emerged as an intermediate cell on the path toward aneuploidy. Approaches that prevent the formation of and/or target existing polyploid cancer cells are actively being investigated. However, we are just beginning to understand how to best attack polyploid cancer cells. It appears combination therapies that attack all cancer cells, but due to unique vulnerabilities can preferential impact polyploid cells, may have promise. Enabling lethal autophagy has emerged as one means to attack the polyploid cell population of cancer. Additional research is also required to investigate the role that polyploid cancer cells could play in anti-tumor immunity. The ER stress response and calreticulin play a role in immune surveillance for aberrant polyploidy, but how this is bypassed to enable the persistence of polyploid cancer cells in patients is enigmatic. Therapeutics that reestablish immune attack on polyploid cells, alongside therapies that preferentially attack the vulnerabilities of polyploids, may prove a potent combination that halts tumor progression in its tracks.

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References


