

Supplementary Materials

Supplementary Figures

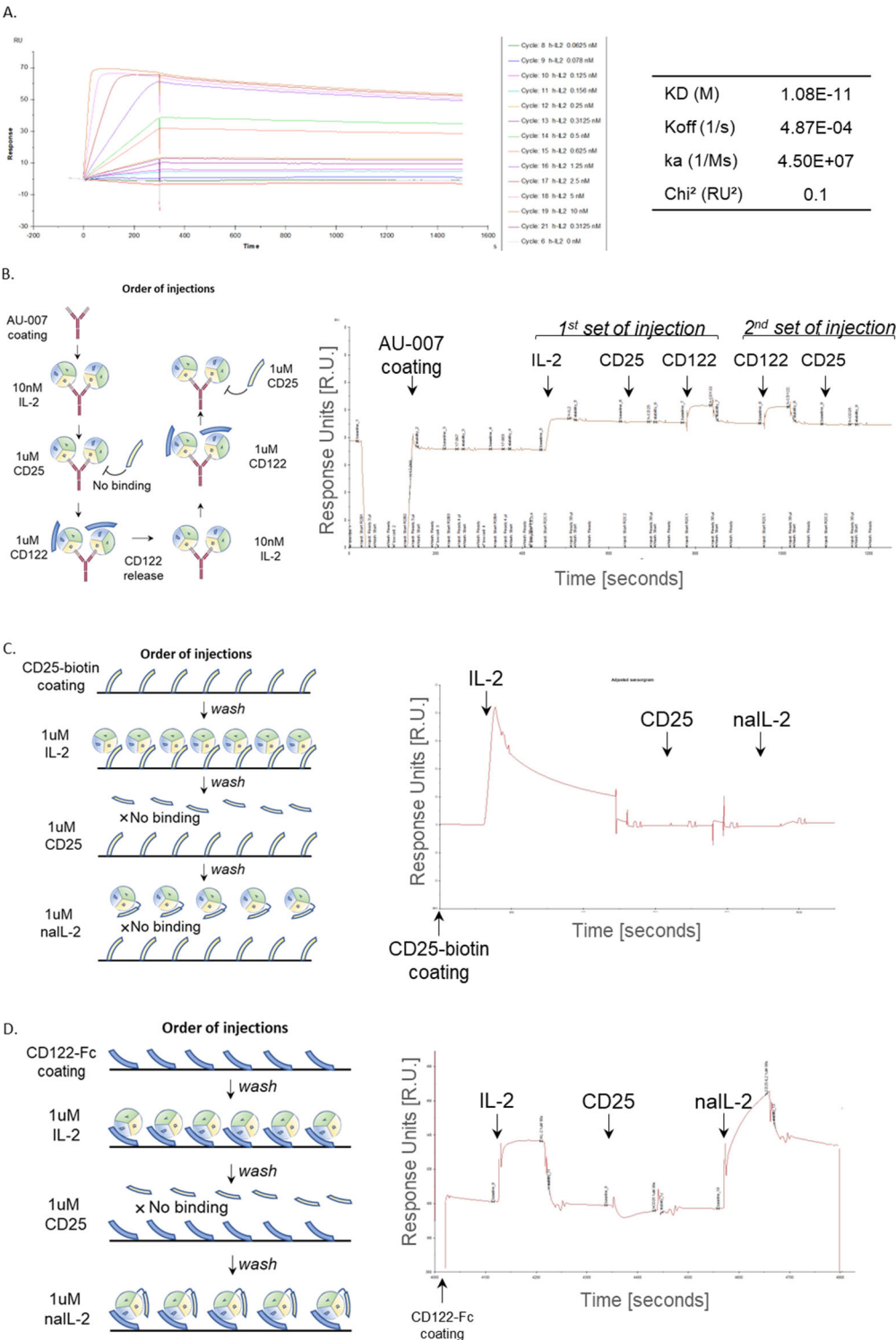


Figure S1. A-B: AU-007 binds human IL-2 with high affinity and inhibits the binding to hCD25 while preserving the binding to hCD122. Affinity and epitope binding site was assessed using Surface Plasmon Resonance (SPR). A. SPR sensorgram traces and calculated binding kinetics of a CM5 chip-bound AU-007 with hIL-2 serving as analyte. B. AU-007 Epitope binding analysis: AU-007 was captured on a CM5 chip and soluble hIL-2 was injected, forming a complex. Subsequently, soluble hCD25 was injected followed by the injection of soluble hCD122. Diagrams represent SPR sensorgram traces of complex formation of Ab/IL-2/IL-2R. Arrows indicate where hIL-2, hCD25, and hCD122 were injected. **C-D:** nIL-2 (*hIL-2/hCD25 conjugate*) inhibits the binding to hCD25 while preserving the binding to hCD122. C. Biotinylated hCD25 was captured on a CM5 chip and soluble hIL-2, soluble hCD25, and nIL-1 were injected subsequently. D. Fc tagged hCD122 was captured on a CM5 chip and soluble hIL-2, soluble hCD25, and soluble nIL-2 were injected subsequently. Diagrams represent SPR sensorgram traces of complex formation of IL-2R/Cytokine. Arrows indicate where hIL-2, hCD25, and hCD122 were injected.

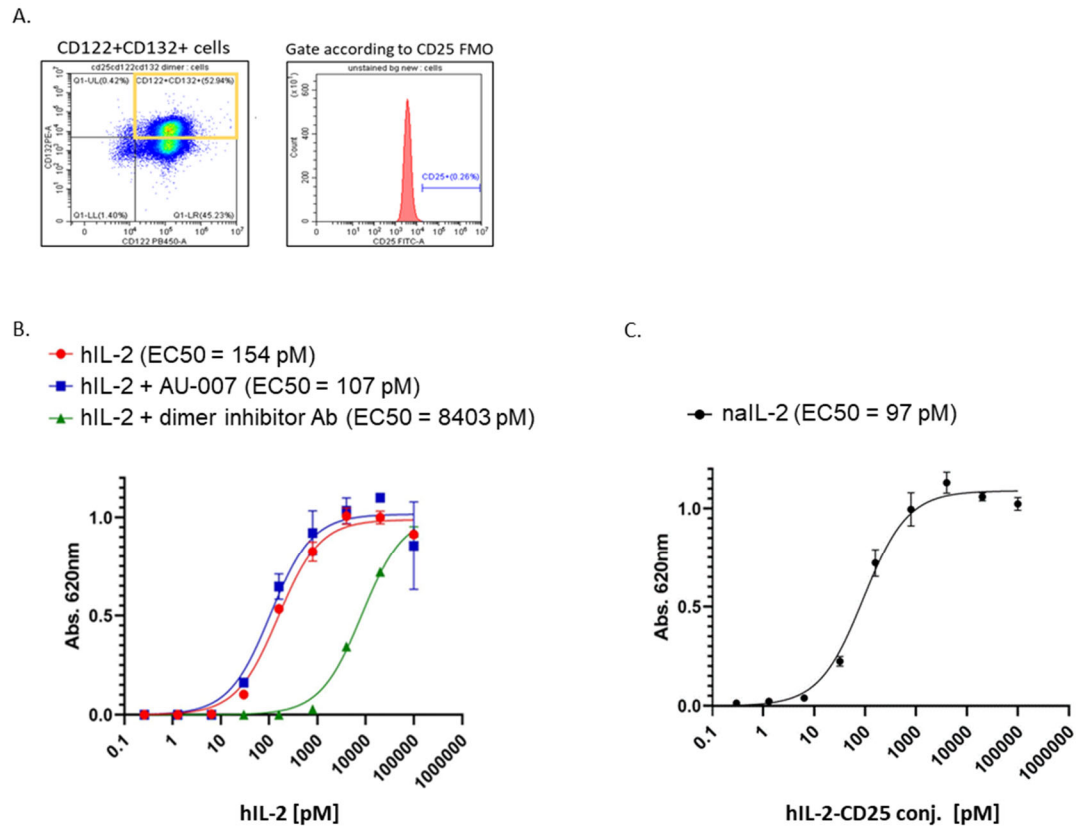


Figure S2. AU-007 and the nall-2 do not hinder CD122/CD132-STAT5 signaling activity. A HEK239-dimer-STAT5-SEAP reporter cell line that stably expresses the human IL-2 dimer receptor (CD122/CD132) with no expression of CD25 and drives the expression of secreted embryonic alkaline phosphatase (SEAP) under a STAT5 promoter, was used to detect IL-2/IL-2-dimeric receptor signaling. A. CD25, CD122, and CD132 expression levels were detected using flow cytometry verifying the exclusive expression of the dimeric receptor. B. Dose response curves of IL-2 alone (red circles) or in the presence of 200nM AU-007 (blue squares) or of 200nM of an anti-IL-2 antibody that inhibits interactions with the dimeric receptor (green triangles). C. Dose response curve of nall-2 (black circles). HEK239-dimer-STAT5-SEAP reporter cells were treated with increasing concentrations of hIL-2 alone or with indicated anti-hIL-2 antibodies (B) or with increasing concentrations of nall-2 (C), 24h post-treatment accumulated levels of SEAP were measured from cells media and functional EC-50 was calculated using GraphPad (B & C).

Supplementary Tables

Table S1. Antibodies used for flow cytometry in vitro experiments of cultured hPBMCs.

Markers	Fluorochrome	Clone	Cat.	Isotypes	Vender
hCD3	PE- CF594	UCHT1	562280	Mouse IgG1, κ	BD
hCD4	FITC	RPA-T4	555346	Mouse IgG1, κ	BD
hCD8	APC-Cy7	RPA-T8	557760	Mouse IgG1, κ	BD
hCD25	BV421	BC96	302630	Mouse IgG1, κ	BioLegend
hCD127	BV711	HIL-7R-M21	563165	Mouse IgG1, κ	BD
hCD56	BV605	HCD56	318334	Mouse IgG1, κ	BioLegend
hFoxP3	PE	150D/E4	12-4774-42	Mouse IgG1, κ	eBioscience
pSTAT5	AF647	47/Stat5(pY694)	562076	Mouse IgG1, κ	BD
Viability dye	EF660	NA	65-0864-18	NA	eBioscience

Table S2. Antibodies used for flow cytometry experiments of hPBMCs that were isolated from spleens of NOG-EXL mice engrafted with hPBMCs.

Markers	Fluorochrome	Clone	Cat.	Isotypes	Vender
mCD45	PerCP-Cy5.5	30-F11	103132	Rat IgG2b	BioLegend
hCD45	BV785	HI30	304048	Mouse IgG1, κ	Biolegend
hCD3	BUV395	SK7	564001	Mouse BALB/c	BD
hCD4	FITC	OKT4	317408	Mouse IgG2b, κ	BioLegend
hCD25	BV711	M-A251	356138	Mouse IgG1, κ	BioLegend
hCD127	PE-Cy7	HIL-7R-M21	560822	Mouse IgG1, κ	BD
hFoxP3*	PE	259D/C7	560082	Mouse BALB/c	BD
hCD8	PE/Dazzle594	SK1	344744	Mouse IgG1, κ	BioLegend
hCD45RA	BV421	HI100	304130	Mouse IgG2b, κ	BioLegend
hCCR7	APC	G043H7	353214	Mouse IgG2a, κ	BioLegend
hCD56	BV605	HCD56	318334	Mouse IgG1, κ	BioLegend
Viability dye	eF780	NA	65-0865-18	NA	eBioscience