

Susceptibility of Malignant Brain Tumors to 5-aminolaevulinic Acid Mediated Photodynamic Therapy: Direct Phototoxicity and Immunological Effects

Angeliki Datsi, Rüdiger V. Sorg*

Institute for Transplantation Diagnostics and Cell Therapeutics, Heinrich-Heine University Hospital, Medical Faculty, Moorenstrasse 5, 40225 Düsseldorf, Germany

*Correspondence should be addressed to Rüdiger V. Sorg, ruediger.sorg@med.uni-duesseldorf.de

Received date: October 31, 2020, **Accepted date:** November 13, 2020

Copyright: © 2020 Datsi A, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Commentary

Recently we published the article ‘Accumulation of protoporphyrin IX in medulloblastoma cell lines and sensitivity to subsequent photodynamic treatment’ [1]. In this commentary, we review protoporphyrin IX accumulation after application of 5-aminolaevulinic acid and the resulting sensitivity of medulloblastoma cells to photodynamic therapy. We compare the results to glioblastoma cells, including glioblastoma stem-like cells, and address the contribution of the transporter adenosine triphosphate binding cassette subfamily G member 2 (ABCG2) as well as the enzyme ferrochelatase to the process. We discuss possible strategies to improve efficiency of photodynamic therapy, particularly in the clinical setting and highlight the contribution of the anti-tumoral immune response to the efficacy of this novel treatment modality for brain tumors.

Novel Therapeutic Options for Malignant Brain Tumors

Primary brain tumors are a heterogeneous group of tumors arising within the central nervous system. They can be benign or malignant and develop in children as well as in adults. The most frequent malignant brain tumor in adults is the glioblastoma (GBM), a grade IV glioma of astrocytic lineage with a yearly incidence of 3 – 4 per 100,000 [2]. Despite multimodal therapy, consisting of maximal safe resection combined with radiotherapy and alkylating chemotherapy with temozolomide, prognosis of patients is poor: median overall survival (mOS) is 15 months, and after three years, virtually all patients have

relapsed with a mOS after recurrence of 6 months [3,4].

In children, GBM is rare (1 – 2 per million; [5]). Here, medulloblastoma (MB), a neuroepithelial tumor located in the cerebellum and the fourth ventricle is the most common malignant brain tumor, accounting for 20% of all pediatric brain tumors, with incidences ranging from 0.6 – 9 per million, depending on age [6,7]. The current treatment combines surgical resection with craniospinal radiotherapy and chemotherapy. However, due to therapy-associated sequelae, radiation sparing protocols are used in infants and reduced-dose protocols in patients at lower risk, e.g. when gross-total resection of the tumor (< 1.5 cm² residual tumor) has been achieved. Five-year survival ranges from 60 to 95%, depending on subtype and risk group [6,8,9].

In both tumors, GBM and MB, the extent of resection is a strong prognostic factor. However, total or even gross-total resection is frequently a neurosurgical challenge. The infiltrative growth of the tumor into adjacent functional areas, the frequently eloquent locations of the tumor and the difficulties to clearly identify the tumor intraoperatively increase the risk for resection-associated morbidity. Moreover, current prognosis for GBM patients and particularly high-risk MB patients even after gross-total resection and despite the multimodal therapies is poor. Thus, new therapeutic modalities are needed.

Photodiagnosis (PD) and photodynamic therapy (PDT) are novel therapeutic approaches in brain tumors. Both rely on the preferential enrichment of a photosensitizer in the tumor tissue. In photodiagnosis, the photosensitizer

is used to visualize the tumor intraoperatively, whereas in photodynamic therapy, the photosensitizer is excited with light of a defined wavelength, which initiates a photochemical reaction that results in the generation of reactive oxygen species, which kill the tumor cells [10].

Protoporphyrin IX (PPIX) is a well-established photosensitizer. It is an intermediate of the heme biosynthesis pathway, and is normally converted by the enzyme ferrochelatase to heme by addition of a ferrous ion. 5-aminolaevulinic acid (5-ALA) is generated from glycine and succinyl-CoA by ALA-synthase in the initial rate limiting step of heme biosynthesis. When it is added to cells exogenously, it is transported into the cytosol by the peptide transporters PEPT1 and PEPT2 and metabolized to coproporphyrinogen III, which is transported into the mitochondria by the transporter ABCB6 and further metabolized via PPIX to heme [11]. In numerous tumor cells, however, conversion of PPIX to heme is impaired due to different mechanisms, including a lack or down-regulation of ferrochelatase [12]. Hence, non-metabolized PPIX accumulates in the mitochondria and the cytosol of the tumor cells.

GBM cells show reduced expression of ferrochelatase and accumulate PPIX when subjected to exogenous 5-ALA *in vitro* [13]. Oral intake of 5-ALA prior to surgery allows intraoperative identification of the tumor when exposed to light of 400 nm wavelength during fluorescence-guided surgery [14]. Thereby, the extent of tumor resection can safely be increased, which provides a survival benefit for the patients [15,16]. Thus, in GBM application of 5-ALA is a powerful photodiagnostic tool.

PPIX accumulation has also been reported for MB cells *in vitro* [1,17-20], however, *in vivo* results have been contradictory. In several studies on pediatric brain tumors, PPIX fluorescence has been detected during fluorescence-guided surgery in only 4 out of 11 MB patients [21-25], and in only 2 of the patients intraoperative identification of the tumor was possible [26]. Moreover, a patchy rather than homogenous fluorescence of the tumors has been described [22,27], which may indicate the presence of distinct subsets of MB cells differing in PPIX accumulation within the same tumor. Indeed, when we compared PPIX accumulation in MB and GBM cell lines, accumulation was more efficient in the GBM cells and not all of the MB cells became PPIX positive [1], supporting this conclusion.

There was no difference in ferrochelatase expression and activity between GBM and MB cells, which would account for the differences [1]. However, we observed expression of the transporter ABCG2 (CD338) on the surface of MB, but not GBM cells [1]. Thus, MB cells express a protein that transports PPIX out of the cells and, therefore, could counteract PPIX accumulation [28].

Whether ABCG2 expression contributes to the differences and identifies different subsets of MB cells is currently under investigation. However, it is tempting to speculate that inhibition of ABCG2, e.g. by imatinib, verapamil or genestein [11,29-31] may increase the efficiency of PPIX accumulation in MB.

Besides the presence of ABCG2, the efficiency of 5-ALA uptake and its metabolization towards PPIX, the availability of ferrous iron or even the formation of non-fluorescent aggregates can affect PPIX accumulation. *In vivo* there may also be differences in pharmacokinetics of 5-ALA and, particularly, the access to the MB tissue could be limited, e.g. when the blood-brain-barrier is undisturbed or capillary permeability and blood flow are low [32,33].

Interestingly, in GBM high expression levels of ABCG2 have been described in glioma stem-like cells (GSC) [34], thus, a cellular subset within the tumor, which has been proposed to initiate and maintain tumor growth [35,36] and to represent the driving force for disease recurrence [35,37]. In GSC, ABCG2 contributes to multidrug resistance, including resistance to temozolomide [38], but probably also to self-renewal and maintenance of stemness [39]. We reported recently that there are differences between GBM cells and primary GSC lines in PPIX accumulation [40] and could show that blocking ABCG2 in GSC increases PPIX accumulation (Ebbert et al., manuscript in preparation).

Little is known about MB stem-like cells and their accumulation of PPIX. Lee et al. isolated CD133⁺ neuronal stem cells from the cerebellum, which may represent the cells of origin of MB [41], and della Puppa et al. identified CD133⁺ MB cells, which were located in the faintly fluorescent external areas of the tumor, whereas the inner core consisted of more fluorescent tissue [27]. However, from these data no conclusion can be drawn on PPIX accumulation in MB cells versus MB stem-like cells and why in MB not all cells appear to accumulate PPIX.

Exposure of GBM or MB cells, which had been subjected to 5-ALA, to laser light of 635 nm wavelength (PDT) initiates a photochemical reaction and results in the generation of reactive oxygen species, including singlet-oxygen, which ultimately causes death of the tumor cells. Consistent with the reduced efficiency of PPIX accumulation in MB, GBM cells are more sensitive to the phototoxic effect of 5-ALA-mediated PDT [1], and a substantial fraction of MB cells survives the treatment. Indeed, photosensitivity of MB cells appears to be directly dependent on PPIX accumulation, since PPIX accumulation and survival after 5-ALA/PDT treatment show an almost linear relationship. Thus, other factors such as oxygen concentration seem to have a minor influence, and to enhance efficacy of 5-ALA/PDT in MB,

it will be important to improve PPIX accumulation as discussed before or other photosensitizers have to be used, such as hypericin [19] and 5-ALA-hexylester [20].

Even in GBM, which responds to 5-ALA/PDT, clinical efficacy may depend on the treatment targeting GSC, because they represent the driving force for disease recurrence [35,37]. When GSC were subjected to 5-ALA/PDT, a 5-ALA-dose-dependent reduction in viability was observed [40]. However, different GSC preparations differed in sensitivity, which followed a negative exponential regression model, suggesting that it mainly depends on the level of PPIX accumulation, i.e. 5-ALA metabolism to PPIX and the efflux of PPIX. As discussed before, GSC express ABCG2 [34], which transports PPIX out of the cells [28], and indeed, blocking ABCG2 in GSC not only results in an increase of PPIX accumulation, but also in increased sensitivity to 5-ALA/PDT (Ebbert et al., manuscript in preparation). Thus, although early clinical reports on the therapeutic use of 5-ALA/PDT are promising [42-45], efficacy in GBM may depend on combining it with at least an inhibitor of the transporter ABCG2. In addition, more effective illumination by novel devices, which allow a more homogenous and controlled delivery of the laser light to the surgical cavity [46], may result in an increased sensitivity of the tumor cells at a given PPIX accumulation, and could thereby contribute to increase efficacy of the treatment.

Irrespective of the device, however, the phototoxic damage caused by 5-ALA/PDT can extend only up to a depth of about 4 mm in cerebral tissue [47]. Therefore, it will not reach the more distant tumor cells. However, 5-ALA/PDT not only directly kills the tumor cells, but it is well established that it generates pro-inflammatory conditions and contributes to the induction of anti-tumoral T-cell responses, with CD8⁺ T cells as main effectors and a supportive role of CD4⁺ T cells [10]. For glioblastoma cells we could demonstrate that 5-ALA/PDT initiates the afferent phase of adaptive immune responses leading to attraction, antigen-uptake and activation of dendritic cells mediated by the release of the heat-shock protein 70 by the dying cells [48], pre-requisites for the initiation of T cell immunity. Yet, it needs to be elucidated whether this results in cross-presentation of tumor antigens and induction of cytotoxic T-cell-mediated anti-tumoral immunity *in vivo*. The importance of such anti-tumoral immune responses to target particularly GSC has been documented by Pellegatta et al., who showed that using neurosphere-derived GSC as a source of tumor antigens improves efficacy of dendritic cell-based immunotherapy in a GL261 glioma mouse model [49]. Since GSC are sensitive to 5-ALA/PDT, the T-cell responses may also be directed towards the GSC compartment in the tumor, further highlighting the possible therapeutic potential of photodynamic therapy.

In conclusion, the selective accumulation of PPIX after the application of 5-ALA in brain tumors such as glioblastoma and medulloblastoma has the potential to improve the outcome of cytoreductive procedures by allowing to identify the tumor intraoperatively during fluorescence-guided surgery. Furthermore, when PPIX is excited with laser light of 635 nm wavelength, a photochemical reaction is initiated, generating reactive oxygen species, which kill the tumor cells. At least in glioblastoma, the respective cancer stem cells appear to be sensitive to 5-ALA/PDT as well, potentially reducing the risk of disease recurrence. Moreover, 5-ALA/PDT not only kills the tumor cells directly due to the phototoxic effect, but also results in the initiation of an anti-tumoral immune response, by attracting dendritic cells, promoting uptake of dying tumor cells and inducing dendritic cell maturation. This immunological effect is likely to contribute to the clinical responses to 5-ALA/PDT. Thus, 5-ALA and PPIX hold photodiagnostic as well therapeutic potential. Whether this approach can be extended to other brain tumors or enhanced, e.g. by inhibiting the enzyme ferrochelatase or the ABCG2 transporter to increase PPIX accumulation, requires further investigations.

Conflict of Interest

The authors declare no conflict of interest

References

1. Briel-Pump A, Beez T, Ebbert L, Remke M, Weinhold S, Sabel MC, et al. Accumulation of protoporphyrin IX in medulloblastoma cell lines and sensitivity to subsequent photodynamic treatment. *J Photochem Photobiol B.* 2018;189:298-305.
2. Ostrom QT, Bauchet L, Davis FG, Deltour I, Fisher JL, Langer CE, et al. The epidemiology of glioma in adults: a “state of the science” review. *Neuro Oncol.* 2014;16(7):896-913.
3. Stupp R, Hegi ME, Mason WP, van den Bent MJ, Taphoorn MJ, Janzer RC, et al. Effects of radiotherapy with concomitant and adjuvant temozolomide versus radiotherapy alone on survival in glioblastoma in a randomised phase III study: 5-year analysis of the EORTC-NCIC trial. *Lancet Oncol.* 2009;10(5):459-66.
4. Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N Engl J Med.* 2005;352(10):987-96.
5. Fisher JL, Schwartzbaum JA, Wrensch M, Wiemels JL. Epidemiology of brain tumors. *Neurol Clin.* 2007;25(4):867-90.

6. Massimino M, Biassoni V, Gandola L, Garre ML, Gatta G, Giangaspero F, et al. Childhood medulloblastoma. *Crit Rev Oncol Hematol.* 2016;105:35-51.
7. Smoll NR, Drummond KJ. The incidence of medulloblastomas and primitive neuroectodermal tumours in adults and children. *J Clin Neurosci.* 2012;19(11):1541-4.
8. Ramaswamy V, Taylor MD. Medulloblastoma: From Myth to Molecular. *J Clin Oncol.* 2017;35(21):2355-63.
9. Schwalbe EC, Lindsey JC, Nakjang S, Crosier S, Smith AJ, Hicks D, et al. Novel molecular subgroups for clinical classification and outcome prediction in childhood medulloblastoma: a cohort study. *Lancet Oncol.* 2017;18(7):958-71.
10. Castano AP, Mroz P, Hamblin MR. Photodynamic therapy and anti-tumour immunity. *Nat Rev Cancer.* 2006;6(7):535-45.
11. Ishikawa T, Nakagawa H, Hagiya Y, Nonoguchi N, Miyatake S, Kuroiwa T. Key Role of Human ABC Transporter ABCG2 in Photodynamic Therapy and Photodynamic Diagnosis. *Adv Pharmacol Sci.* 2010;2010:587306.
12. Ohgari Y, Nakayasu Y, Kitajima S, Sawamoto M, Mori H, Shimokawa O, et al. Mechanisms involved in delta-aminolevulinic acid (ALA)-induced photosensitivity of tumor cells: relation of ferrochelatase and uptake of ALA to the accumulation of protoporphyrin. *Biochem Pharmacol.* 2005;71(1-2):42-9.
13. Teng L, Nakada M, Zhao SG, Endo Y, Furuyama N, Nambu E, et al. Silencing of ferrochelatase enhances 5-aminolevulinic acid-based fluorescence and photodynamic therapy efficacy. *Br J Cancer.* 2011;104(5):798-807.
14. Stummer W, Stocker S, Wagner S, Stepp H, Fritsch C, Goetz C, et al. Intraoperative detection of malignant gliomas by 5-aminolevulinic acid-induced porphyrin fluorescence. *Neurosurgery.* 1998;42(3):518-25.
15. Stummer W, Reulen HJ, Meinel T, Pichlmeier U, Schumacher W, Tonn JC, et al. Extent of resection and survival in glioblastoma multiforme: identification of and adjustment for bias. *Neurosurgery.* 2008;62(3):564-76.
16. Stummer W, Pichlmeier U, Meinel T, Wiestler OD, Zanella F, Reulen HJ. Fluorescence-guided surgery with 5-aminolevulinic acid for resection of malignant glioma: a randomised controlled multicentre phase III trial. *Lancet Oncol.* 2006;7(5):392-401.
17. Schwake M, Nemes A, Dondrop J, Schroeteler J, Schipmann S, Senner V, et al. In-Vitro Use of 5-ALA for Photodynamic Therapy in Pediatric Brain Tumors. *Neurosurgery.* 2018;83(6):1328-37.
18. Schwake M, Gunes D, Kochling M, Brentrup A, Schroeteler J, Hotfilder M, et al. Kinetics of porphyrin fluorescence accumulation in pediatric brain tumor cells incubated in 5-aminolevulinic acid. *Acta Neurochir (Wien).* 2014;156(6):1077-84.
19. Ritz R, Scheidle C, Noell S, Roser F, Schenk M, Dietz K, et al. In vitro comparison of hypericin and 5-aminolevulinic acid-derived protoporphyrin IX for photodynamic inactivation of medulloblastoma cells. *PLoS One.* 2012;7(12):e51974.
20. Chu ES, Wong TK, Yow CM. Photodynamic effect in medulloblastoma: downregulation of matrix metalloproteinases and human telomerase reverse transcriptase expressions. *Photochem Photobiol Sci.* 2008;7(1):76-83.
21. Skjoth-Rasmussen J, Bogeskov L, Sehested A, Klausen C, Broholm H, Nysom K. The use of 5-ALA to assist complete removal of residual non-enhancing part of childhood medulloblastoma: a case report. *Childs Nerv Syst.* 2015;31(11):2173-7.
22. Preuss M, Renner C, Krupp W, Christiansen H, Fischer L, Merckenschlager A, et al. The use of 5-aminolevulinic acid fluorescence guidance in resection of pediatric brain tumors. *Childs Nerv Syst.* 2013;29(8):1263-7.
23. Eicker S, Sarikaya-Seiwert S, Borkhardt A, Gierga K, Turowski B, Heiroth HJ, et al. ALA-induced porphyrin accumulation in medulloblastoma and its use for fluorescence-guided surgery. *Cent Eur Neurosurg.* 2011;72(2):101-3.
24. Beez T, Sarikaya-Seiwert S, Steiger HJ, Hanggi D. Fluorescence-guided surgery with 5-aminolevulinic acid for resection of brain tumors in children--a technical report. *Acta Neurochir (Wien).* 2014;156(3):597-604.
25. Barbagallo GM, Certo F, Heiss K, Albanese V. 5-ALA fluorescence-assisted surgery in pediatric brain tumors: report of three cases and review of the literature. *Br J Neurosurg.* 2014;28(6):750-4.
26. Stummer W, Rodrigues F, Schucht P, Preuss M, Wiewrodt D, Nestler U, et al. Predicting the "usefulness" of 5-ALA-derived tumor fluorescence for fluorescence-guided resections in pediatric brain tumors: a European survey. *Acta Neurochir (Wien).* 2014;156(12):2315-24.
27. Della Puppa A, Gioffre G, Gardiman MP, Frasson C, Cecchin D, Scienza R, et al. Intra-operative 5-aminolevulinic acid (ALA)-induced fluorescence of medulloblastoma: phenotypic variability and CD133(+)

expression according to different fluorescence patterns. *Neurol Sci.* 2014;35(1):99-102.

28. Nakanishi T, Ogawa T, Yanagihara C, Tamai I. Kinetic Evaluation of Determinant Factors for Cellular Accumulation of Protoporphyrin IX Induced by External 5-Aminolevulinic Acid for Photodynamic Cancer Therapy. *J Pharm Sci.* 2015;104(9):3092-100.

29. Bebes A, Nagy T, Bata-Csorgo Z, Kemeny L, Dobozy A, Szell M. Specific inhibition of the ABCG2 transporter could improve the efficacy of photodynamic therapy. *J Photochem Photobiol B.* 2011;105(2):162-6.

30. Fujita H, Nagakawa K, Kobuchi H, Ogino T, Kondo Y, Inoue K, et al. Phytoestrogen Suppresses Efflux of the Diagnostic Marker Protoporphyrin IX in Lung Carcinoma. *Cancer Res.* 2016;76(7):1837-46.

31. Ogino T, Kobuchi H, Munetomo K, Fujita H, Yamamoto M, Utsumi T, et al. Serum-dependent export of protoporphyrin IX by ATP-binding cassette transporter G2 in T24 cells. *Mol Cell Biochem.* 2011;358(1-2):297-307.

32. Warnke PC, Kopitzki K, Timmer J, Ostertag CB. Capillary physiology of human medulloblastoma: impact on chemotherapy. *Cancer.* 2006;107(9):2223-7.

33. Wolburg H, Noell S, Fallier-Becker P, Mack AF, Wolburg-Buchholz K. The disturbed blood-brain barrier in human glioblastoma. *Mol Aspects Med.* 2012;33(5-6):579-89.

34. Chen L, Shi L, Wang W, Zhou Y. ABCG2 downregulation in glioma stem cells enhances the therapeutic efficacy of demethoxycurcumin. *Oncotarget.* 2017;8(26):43237-47.

35. Lathia JD, Mack SC, Mulkearns-Hubert EE, Valentim CL, Rich JN. Cancer stem cells in glioblastoma. *Genes Dev.* 2015;29(12):1203-17.

36. Vescovi AL, Galli R, Reynolds BA. Brain tumour stem cells. *Nat Rev Cancer.* 2006;6(6):425-36.

37. Auffinger B, Tobias AL, Han Y, Lee G, Guo D, Dey M, et al. Conversion of differentiated cancer cells into cancer stem-like cells in a glioblastoma model after primary chemotherapy. *Cell Death Differ.* 2014;21(7):1119-31.

38. Eramo A, Ricci-Vitiani L, Zeuner A, Pallini R, Lotti F, Sette G, et al. Chemotherapy resistance of glioblastoma stem cells. *Cell Death Differ.* 2006;13(7):1238-41.

39. Susanto J, Lin YH, Chen YN, Shen CR, Yan YT, Tsai ST, et al. Porphyrin homeostasis maintained by ABCG2 regulates self-renewal of embryonic stem cells. *PLoS One.* 2008;3(12):e4023.

40. Schimanski A, Ebbert L, Sabel MC, Finocchiaro G, Lamszus K, Ewelt C, et al. Human glioblastoma stem-like cells accumulate protoporphyrin IX when subjected to exogenous 5-aminolaevulinic acid, rendering them sensitive to photodynamic treatment. *Journal of photochemistry and photobiology B, Biology.* 2016;163:203-10.

41. Lee A, Kessler JD, Read TA, Kaiser C, Corbeil D, Huttner WB, et al. Isolation of neural stem cells from the postnatal cerebellum. *Nat Neurosci.* 2005;8(6):723-9.

42. Beck TJ, Kreth FW, Beyer W, Mehrkens JH, Obermeier A, Stepp H, et al. Interstitial photodynamic therapy of nonresectable malignant glioma recurrences using 5-aminolevulinic acid induced protoporphyrin IX. *Lasers Surg Med.* 2007;39(5):386-93.

43. Eljamel MS, Goodman C, Moseley H. ALA and Photofrin fluorescence-guided resection and repetitive PDT in glioblastoma multiforme: a single centre Phase III randomised controlled trial. *Lasers Med Sci.* 2008;23(4):361-7.

44. Stummer W, Beck T, Beyer W, Mehrkens JH, Obermeier A, Etminan N, et al. Long-sustaining response in a patient with non-resectable, distant recurrence of glioblastoma multiforme treated by interstitial photodynamic therapy using 5-ALA: case report. *J Neurooncol.* 2008;87(1):103-9.

45. Johansson A, Faber F, Kniebuhler G, Stepp H, Sroka R, Egensperger R, et al. Protoporphyrin IX fluorescence and photobleaching during interstitial photodynamic therapy of malignant gliomas for early treatment prognosis. *Lasers Surg Med.* 2013;45(4):225-34.

46. Dupont C, Vermandel M, Leroy HA, Quidet M, Lecomte F, Delhem N, et al. INtraoperative photoDYNAMIC Therapy for GliOblastomas (INDYGO): Study Protocol for a Phase I Clinical Trial. *Neurosurgery.* 2019;84(6):E414-E9.

47. Olzowy B, Hundt CS, Stocker S, Bise K, Reulen HJ, Stummer W. Photoirradiation therapy of experimental malignant glioma with 5-aminolevulinic acid. *J Neurosurg.* 2002;97(4):970-6.

48. Etminan N, Peters C, Lakbir D, Bunemann E, Borger V, Sabel MC, et al. Heat-shock protein 70-dependent dendritic cell activation by 5-aminolevulinic acid-mediated photodynamic treatment of human glioblastoma spheroids in vitro. *Br J Cancer.* 2011;105(7):961-9.

49. Pellegatta S, Poliani PL, Corno D, Menghi F, Ghielmetti F, Suarez-Merino B, et al. Neurospheres enriched in cancer stem-like cells are highly effective in eliciting a dendritic cell-mediated immune response against malignant gliomas. *Cancer Res.* 2006;66(21):10247-52.