

Obeticholic Acid, FXR Agonists, Liver Disease and Plasma Biomarkers

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Abstract

Although it is recognized that more biomarkers are needed to evaluate the progression of liver disease and response to medications, plasma alkaline phosphatase and conjugated bilirubin are a focus of clinical trials and utilized by physicians in practice. Conjugated bilirubin is a surrogate marker for the status of bile acid transport and the lowering of plasma levels in response to administration of ursodeoxycholic acid indicates that a choleric effect has occurred. Plasma alkaline phosphatase levels are affected by hepatic bile acid composition and flow and the status of the cholangioles. Medications that reduce endogenous bile acid pool size can augment the effect of ursodeoxycholic acid by expanding its proportion in those undergoing enterohepatic circulation. In addition, they can lower the elevated steady state concentration of hepatocellular bile acids and retard progression to cirrhosis.

Keywords: Plasma biomarkers, Conjugated bilirubin, Alkaline phosphatase, FXR agonists, Ursodeoxycholic acid, Obeticholic acid, Cholangioles, Canalicular conduit

Abbreviations: BSEP: Bile Salt Excretory Pump; NTCP: Sodium/ bile acid cotransporter; FXR: Farnesoid X Receptor

Introduction

How medications whose major biologic effect is to reduce bile acid synthesis favorably affect the course of a variety of cholestatic and metabolic liver diseases is not immediately apparent. Also, the most frequently used plasma biomarkers for evaluating benefit, alkaline phosphatase and conjugated bilirubin [1], provide different information. The former may be misleading with respect to the course of the disease and therefore it is important to focus on the pathophysiologic basis for its use.

Hepatocellular Concentration of Bile Acids and Liver Disease

Two mutations in bile acid transporters NTCP and BSEP, which render them non-functional, indicate the importance of the hepatocellular concentration of bile acids. The initial report of a mutation in NTCP was in a child with elevated plasma bile acids since birth, no liver disease and, perhaps surprisingly, no pruritus [2]. By

contrast, mutations in BSEP (PFIC2) [3] always lead to cirrhosis early in life. Since NTCP regulates entry of bile acids into the hepatocyte and BSEP regulates their exit, it is reasonable to extrapolate from this knowledge that an increase in their hepatocellular concentration sets in motion a variety of processes leading to cirrhosis [4].

Further strong support for this view is the identification of families with non-functional mutations in FXR with severe cholestatic liver disease [5]. The characteristic plasma biomarkers are elevations in the concentration of conjugated bilirubin and bile acids but normal gamma-glutamyl transferase levels, regarded as a more sensitive and precise surrogate for alkaline phosphatase.

A biomarker or technique that provides a precise knowledge of the hepatocellular concentration of bile acids would be the most useful test for managing patients with liver disease, but none exists. A technique utilizing [*N*-methyl-¹¹C] choly sarcosine (11S-CSar), a surrogate for endogenous bile acids, does permit visualization of the

hepatocellular transport system and can be informative but requires PET scanning and currently is of relatively little clinical use [6]. Also, without knowledge of changes in total bile acid pool size, the information can be misleading [7].

Thus, the current state of the art for both physicians and regulatory agencies focuses mostly on changes in plasma alkaline phosphatase and/or gamma glutamyl transferase and conjugated bilirubin [1].

Plasma Alkaline Phosphatase and Cholestasis

The beneficial effect of ursodeoxycholic acid in treating primary biliary cholangitis (PBC) was established by a prompt fall in the concentration of both plasma alkaline phosphatase and conjugated bilirubin [8]. By contrast, obeticholic acid, when used alone, only led to initially a significant reduction in plasma alkaline phosphatase [9]. Focusing on the differences in the changes in these plasma biomarkers provides insight into their biologic effects.

The normal level of plasma alkaline phosphatase derived from the liver represents a fraction of the protein that entered bile at the canalicular level and re-entered plasma via the sinusoidal blood and hepatic vein as the fluid flowed through the cholangioles. Thus, it is known that the protein is present in the canalicular membrane [10] but absent in cholangioles [11] and that acute bile duct obstruction in the rat leads to a progressive rise in plasma alkaline phosphatase concentration without a change in the concentration of the protein in thoracic duct lymph [12]. It is also known that in humans with an elevated plasma alkaline phosphatase concentration attributable to liver disease the level in thoracic duct lymph is unchanged from normal [13]. The fraction that is returned to plasma via the cholangioles reflects their integrity.

Although it is recognized that plasma alkaline phosphatase without further differentiation is a composite of proteins derived from different organs [14] and that plasma gamma glutamyl transferase, generally considered a more selective and sensitive surrogate for alkaline phosphatase, is also derived from heterogeneous sources [15], routine analyses do not make these differentiations and probably account for the variability sometime found in attempting to correlate these two parameters.

The amount of alkaline phosphatase that enters the canaliculi is governed by the rate of secretion of endogenous bile acids and their generation of cholesterol-lecithin vesicles in which the protein is found as a component [16]. Ursodeoxycholic acid transported into the canalicular conduit mostly as glycine and taurine conjugates generates

a higher flow rate (choleretic) than the endogenous bile acids and reduces micellar concentration [17]. Thus, alkaline phosphatase excretion in bile is less than that of equimolar amounts of endogenous bile acids [17]. Less delivery of alkaline phosphatase to the cholangioles can account for the lowering of plasma levels.

Obeticholic acid, in contrast to ursodeoxycholic acid, is a potent agonist of FXR [18], which downregulates the production of cholesterol 7 alpha hydroxylase, the rate-limiting enzyme in the major pathway of bile acid synthesis. Reducing endogenous pool size leads to reducing the amount of conjugated bile acid transported into the canalicular system during a 24 hr period and thus also to a reduction in alkaline phosphatase secretion. Lowering the synthesis of bile acids within the hepatocyte can lower their elevated intracellular levels, which could be further enhanced by up-regulating BSEP expression.

The choleretic effect of obeticholic acid described in bile duct-cannulated animals [19] when given in doses comparable to chenodeoxycholic acid from which it is derived, is not applicable to the much lower doses administered to humans.

Obeticholic acid when given in combination with ursodeoxycholic acid has the potential for enhancing the excretion of the conjugates of ursodeoxycholic acid by lowering endogenous bile acid pool size and thus expanding its proportion in the circulating pool of bile acids, which will generate a greater canalicular water flow.

The risks that occur in the prolonged administration of ursodeoxycholic acid are beyond the scope of this report. The recent report by Kotb [20] of the increased mortality in a large group of infants and children receiving ursodeoxycholic acid for liver disease compared with a control group should alert physicians to currently unaddressed problems with its prolonged use.

The FXR agonist cilofexor causes a significant reduction of gamma glutamyl transferase in patients with primary sclerosing cholangitis with no significant change in total plasma bilirubin [21]. A significant reduction in secondary bile acids also occurred, which correlates with the reduction in bile acid pool size with a resultant increase in the proportion of bile acids that undergo ileal transport and decrease in bile acid entering the colon.

Fenofibric acid, the active drug produced in the intestines from the ester pro-drug fenofibrate [21] lowers the activity of cholesterol 7 alpha hydroxylase, thus reducing the pool size of endogenous bile acids. Interest in its use for the treatment of PBC arose when fibrates were found to lower plasma alkaline phosphatase and gamma glutamyl transferase activity in individuals who did not have liver

disease [22]. Because of its effect on endogenous bile acid pool size, it can enhance the therapeutic effect of ursodeoxycholic acid in the treatment of PBC [23].

Decreasing the bile acid pool size in treating cholestatic liver disease and metabolic liver disease, when there is evidence for reduced bile acid transport, has as its goal the lowering of the hepatocellular concentration of bile acids. It is reasonable to think that reducing bile acid pool size will lead to a reduction in the mean hepatocellular concentration of bile acids over a 24 hr period. However, considerable individual variation in response may occur depending on the status of NTCP and BSEP transporters. Also, a severe, unintended reduction in bile acid pool size can compromise hepatic bile formation leading to toxicity [24]. Thus, a method for specifically evaluating bile acid transport before and during therapy will provide more precise management of this aspect of the liver disease.

Plasma Conjugated Bilirubin and Cholestasis: A Surrogate Marker for Bile Acid Excretion and Canalicular Water Flow

It is the differences in bile flow that occur with obeticholic acid /or cilofexor and ursodeoxycholic acid that accounts for the lowering of conjugated bilirubin only with the latter. A linkage between the rate of excretion of organic anions and bile acid excretion and canalicular bile flow was demonstrated in the seminal studies of O'Maille [25-28]. Essentially, it was found that the capacity to excrete BSP (Bromsulphalein), which shares the same transport carriers as conjugated bilirubin, is increased by increasing bile acid-dependent canalicular flow but not enhanced by secretin administration, which increases flow from the cholangioles.

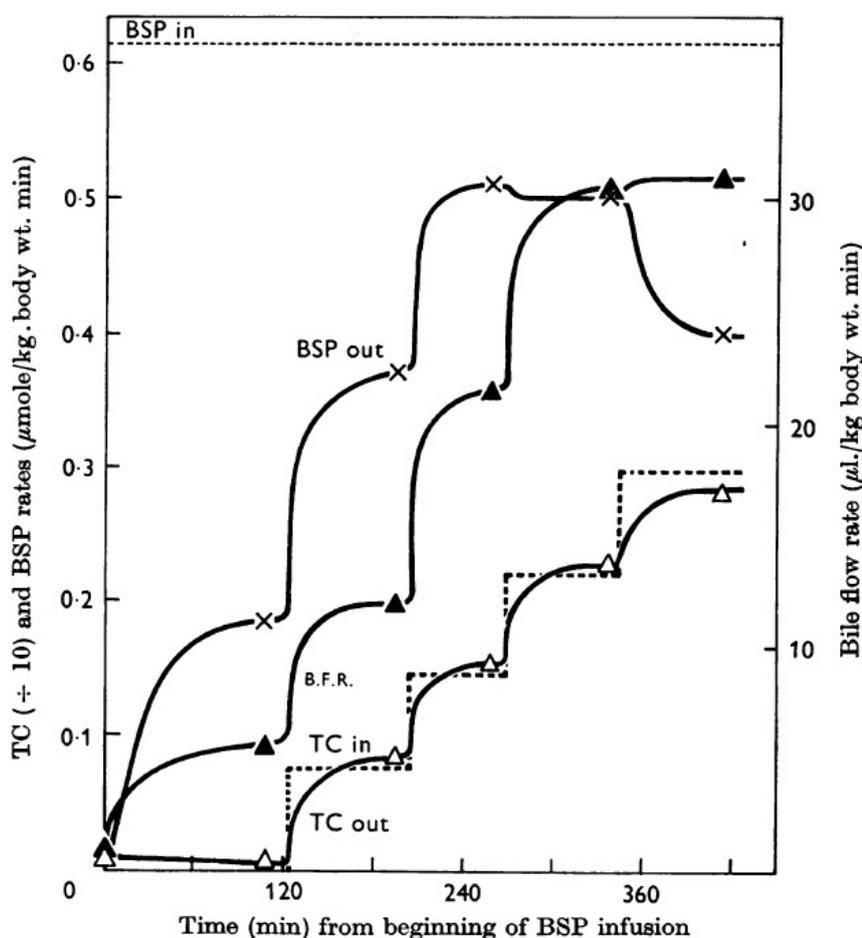


Figure 1: Effect of sodium taurocholate on organic anion excretion. The apparent maximum rate of excretion of an organic anion, phenoltetrabromphthalein disulfonate (Bromsulphalein) is increased (BSP out) when bile flow rate (B.F.R.) is increased by the infusion of sodium taurocholate (TC). From O'Maille, Richards, and Short Reference # 27.

Figure 1 from O'Maille's studies illustrates the effect of increasing bile acid-dependent canalicular bile flow on the biliary excretion of the organic anion, Bromsulphalein. Because on a molar basis the conjugates of ursodeoxycholic acid will increase canalicular bile flow to a greater extent than sodium taurocholate, an increase in conjugated bilirubin excretion and lowering of plasma levels will occur, good evidence for a beneficial effect of therapy.

In using conjugated bilirubin as a surrogate marker of a choleric effect, a limitation can be a change in renal function that can also affect the plasma level.

Author Contribution Statement

Norman B Javitt assembled the subject matter and wrote the entire manuscript.

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References

1. Lammers WJ, Van Buuren HR, Hirschfield GM, Janssen HL, Invernizzi P, Mason AL, et al. Levels of alkaline phosphatase and bilirubin are surrogate end points of outcomes of patients with primary biliary cirrhosis: an international follow-up study. *Gastroenterology*. 2014 Dec 1;147(6):1338-49.

2. Vaz FM, Paulusma CC, Huidekoper H, de Ru M, Lim C, Koster J, et al. Sodium taurocholate cotransporting polypeptide (SLC10A1) deficiency: conjugated hypercholanemia without a clear clinical phenotype. *Hepatology*. 2015 Jan;61(1):260-7.

3. Pawlikowska L, Strautnieks S, Jankowska I, Czubkowski P, Emerick K, Antoniou A, et al. Differences in presentation and progression between severe FIC1 and BSEP deficiencies. *Journal of Hepatology*. 2010 Jul 1;53(1):170-8.

4. Chow MD, Lee YH, Guo GL. The role of bile acids in nonalcoholic fatty liver disease and nonalcoholic steatohepatitis. *Molecular Aspects of Medicine*. 2017 Aug 1;56:34-44.

5. Gomez-Ospina N, Potter CJ, Xiao R, Manickam K, Kim MS, Kim KH, et al. Mutations in the nuclear bile acid receptor FXR cause progressive familial intrahepatic cholestasis. *Nature Communications*. 2016 Feb 18;7(1):1-8.

6. Kjærgaard K, Frisch K, Sørensen M, Munk OL,

Hofmann AF, Horsager J, et al. Obeticholic acid improves hepatic bile acid excretion in patients with primary biliary cholangitis. *Journal of Hepatology*. 2021 Jan 1;74(1):58-65.

7. Javitt NB. Obeticholic acid and hepatic bile acids: Excellent study faulty conclusion. *Journal of Hepatology*. 2021 May 1;74(5):1267.

8. Poupon R, Poupon R, Calmus Y, Chrétien Y, Ballet F, Darnis F. Is ursodeoxycholic acid an effective treatment for primary biliary cirrhosis?. *The Lancet*. 1987 Apr 11;329(8537):834-6.

9. Kowdley KV, Luketic V, Chapman R, Hirschfield GM, Poupon R, Schramm C, et al. A randomized trial of obeticholic acid monotherapy in patients with primary biliary cholangitis. *Hepatology*. 2018 May;67(5):1890-902.

10. Nevens F, Andreone P, Mazzella G, Strasser SI, Bowlus C, Invernizzi P, et al. A placebo-controlled trial of obeticholic acid in primary biliary cholangitis. *New England Journal of Medicine*. 2016 Aug 18;375(7):631-43.

11. Sampaziotis F, Muraro D, Tysoe OC, Sawiak S, Beach TE, Godfrey EM, et al. Cholangiocyte organoids can repair bile ducts after transplantation in the human liver. *Science*. 2021 Feb 19;371(6531):839-46.

12. Baker AL, Hauser SC. Alkaline Phosphatase Activity in Lymph and Serum of Bile Duct-Ligated Rats. *Digestion*. 1978;18(1-2):103-9.

13. Brzek V. Composition of lymph from the thoracic duct in patients with extrahepatic biliary obstruction (author's transl). *Ceskoslovenska Gastroenterologie a Vyziva*. 1975 Mar;29(2):84-8.

14. Korner NH. Distribution of alkaline phosphatase in serum protein fractions. *Journal Of Clinical Pathology*. 1962 May 1;15(3):195-9.

15. Elawdi HA, Franzini M, Paolicchi A, Emdin M, Fornaciari I, Fierabracci V, et al. Circulating gamma-glutamyltransferase fractions in cirrhosis. *Liver International*. 2014 Aug;34(7):e191-9.

16. Crawford JM, Möckel GM, Crawford AR, Hagen SJ, Hatch VC, Barnes S, et al. Imaging biliary lipid secretion in the rat: ultrastructural evidence for vesiculation of the hepatocyte canalicular membrane. *Journal of Lipid Research*. 1996 Jan 1;36(10):2147-63.

17. Hatoff DE, Hardison WG. Bile acid-dependent secretion of alkaline phosphatase in rat bile. *Hepatology*. 1982 Jul;2(4):433S-9S.

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18. Fiorucci S, Cipriani S, Mencarelli A, Baldelli F, Bifulco G, Zampella A. Farnesoid X receptor agonist for the treatment of liver and metabolic disorders: focus on 6-ethyl-CDCA. *Mini reviews in medicinal chemistry.* 2011 Aug 1;11(9):753-62.
 19. Fiorucci S, Clerici C, Antonelli E, Orlandi S, Goodwin B, Sadeghpour BM, et al. Protective effects of 6-ethyl chenodeoxycholic acid, a farnesoid X receptor ligand, in estrogen-induced cholestasis. *Journal of Pharmacology and Experimental Therapeutics.* 2005 May 1;313(2):604-12.
 20. Kotb MA, Mosallam D, Basanti CW, El Sorogy ST, Badr AM, Abd El Baky HE, et al. Ursodeoxycholic acid use is associated with significant risk of morbidity and mortality in infants with cholestasis: a strobe compliant study. *Medicine.* 2020 Feb;99 e18730.
 21. Trauner M, Gulamhusein A, Hameed B, Caldwell S, Shiffman ML, Landis C, et al. The nonsteroidal farnesoid X receptor agonist cilofexor (GS-9674) improves markers of cholestasis and liver injury in patients with primary sclerosing cholangitis. *Hepatology.* 2019 Sep;70(3):788-801.
 22. Day AP, Feher MD, Chopra R, Mayne PD. The effect of bezafibrate treatment on serum alkaline phosphatase isoenzyme activities. *Metabolism.* 1993 Jul 1;42(7):839-42.
 23. Kanda T, Yokosuka O, Imazeki F, Saisho H. Bezafibrate treatment: a new medical approach for PBC patients?. *Journal of Gastroenterology.* 2003 Jun 1;38(6):573-8.
 24. Fiorucci S, Di Giorgio C, Distrutti E. Obeticholic acid: an update of its pharmacological activities in liver disorders. *Bile Acids and Their Receptors.* 2019:283-95.
 25. O'Máille ER. The influence of micelle formation on bile salt secretion. *The Journal of Physiology.* 1980 May 1;302(1):107-20.
 26. O'Máille ER, Richards TG. Possible explanations for the differences in secretory characteristics between conjugated and free bile acids. *The Journal of Physiology.* 1977 Mar 1;265(3):855-66.
 27. o'Maille ER, Richards TG, Short AH. Factors determining the maximal rate of organic anion secretion by the liver and further evidence on the hepatic site of action of the hormone secretin. *The Journal of Physiology.* 1966 Oct 1;186(2):424-38.
 28. O'Máille ER, Richards TG, Short AH. Observations on the elimination rates of single injections of taurocholate and cholate in the dog. *Quarterly Journal of Experimental Physiology and Cognate Medical Sciences: Translation and Integration.* 1969 Jul 16;54(3):296-310.