

Altered Peak C-peptide and Fasting Blood Glucose in Children with Autism Spectrum Disorder

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Abstract

Autism spectrum disorder (ASD) is a series of complex neurobiological disorders. Adolescents and young adults with ASD are more likely to develop type 2 diabetes mellitus in their later life than those who are not. However, whether ASD patients exhibited a pre-diabetic phenotype in their childhood is still unknown. To answer this question, here we presented this study which focus on whether oral glucose tolerance, insulin and C-peptide release were changed in children with ASD. We recruited 17 patients and 17 age and sex-matched typically developing children in our study. In our study, the fasting blood glucose was significantly decreased in children with ASD. Moreover, two children in the ASD group had impaired glucose tolerance according to 2-hour postprandial blood glucose level. The C-peptide level at 30 minutes after glucose administration in the ASD group was significantly lower than that in the control group. Moreover, four children in the ASD group showed a phenomenon of delayed insulin secretion. All these results suggested that a subgroup of children with ASD might exhibit a pre-diabetic phenotype during early childhood. Finally, this study might bring about new therapeutic interventions for children with ASD.

Lay Summary: The etiology of ASD seems to involve an intricate interplay of genetic, environmental and metabolic disorders. Adolescents and young adults with ASD are more likely to develop type 2 diabetes mellitus in their later life. It is important to find out whether ASD patients exhibited a pre-diabetic phenotype in their childhood. This study provided evidence for dysfunction risk of glucose metabolism in ASD and revealed a pre-diabetes phenotype in children with ASD.

Keywords: ASD, Blood glucose, Insulin, C-peptide, IGT

Introduction

ASD refers to a group of complex neurodevelopmental disorders, characterized by deficits in social communication, interaction and demonstrating restricted, repetitive, and stereotyped patterns of behavior. Centers for Disease Control and Prevention forecasted that the prevalence of ASD would be 1 in 45 in USA [1]. A meta-

analysis of the public health and primary care centers in the UK estimated that the prevalence of ASD was 26.6 per 10,000 in mainland, based on eighteen epidemiological studies [2]. Vast clinical heterogeneity is an important feature of ASD. ASD exists tremendous phenotypic heterogeneity in neurological comorbidities, language and cognitive abilities, and adaptive function [3]. ASD patients often have co-occurring seizures, sleep problems,

metabolic disorders and gastrointestinal (GI) disorders [4] Some children with ASD suffer from metabolic problems, including abnormal tyrosine, phenylalanine, tryptophan, asparagine and arginine metabolic pathways [5] Many factors have been demonstrated to be involved in the etiology of ASD, including gut microbiota imbalance [6-8], increased inflammation [9,10], immune system dysfunction [11,12] and altered metabolic capacity [13,14]. Thus, characterizing biological subtypes of ASD will play a key role in diagnosis and treatment.

Nowadays, diabetes mellitus (DM) becomes a global health problem. A higher rate of obesity, impaired glucose tolerance (IGT), cardiovascular and metabolic disorders would increase the risk of neurobehavioral disorders and intellectual disability [15]. Newborns from mothers with gestational diabetes mellitus (GDM) tend to exhibit impaired motor functions, hyperactivity, inattention, and lower intelligence quotient (IQ) scores [16,17]. Type 2 diabetes mellitus (T2DM) can affect cognition and increase dementia risks [18]. Individuals with T2DM during experimental hyperglycemia presented impairments in cognitive and mood state performance [19]. Maternal history of type 1 diabetes mellitus (T1DM) is associated with increased risk of infantile autism in offspring [20]. Another study showed that GDM was associated with the risk of ASD in offspring [21]. Besides, a longitudinal study in Taiwan discovered that adolescents and young adults with ASD were more likely to develop T2DM in their later life [22]. In addition, obesity and diabetes were more common among mothers of children with ASD compared with controls [23]. Compared with typically developing children, the relative risk of overweight and obesity in children with ASD was 2.24 and 4.83, respectively [24]. Obesity and overweight can also increase the prevalence of T2DM and the low-grade inflammation caused by obesity can disturb insulin function [25,26].

Insulin resistance and β -cell dysfunction are two development processes of T2DM. Insulin resistance can lead to an increased demand for insulin. With the development of insulin resistance, insulin is insufficient to meet the requirement, and then β -cell dysfunction will emerge. C-peptide is a better measure of portal insulin secretion than insulin itself [27]. Some studies have reported that young adults with ASD were more likely to develop T2DM in their later life. These young adults with ASD may show prediabetes before they are diagnosed as T2DM. Prediabetes is the precursor stage before DM. It is thought to be a risk state, with high possibilities of developing DM. Impaired fasting blood sugar and IGT are two forms of prediabetes. Early diagnosis and intervention can cure prediabetes and prevent the development of DM [28,29].

However, researches about the association between DM and ASD mainly focused on maternal diabetes. Few studies have focused on glucose metabolism, insulin and C-peptide secretion in children with ASD. Whether precursor symptoms of DM such as insulin resistance and IGT occurred in their childhood are still unknown. To answer this question, we performed oral glucose tolerance test (OGTT), insulin and C peptide release tests in children with ASD and the control children. The results showed that fasting blood glucose significantly decreased in children with ASD and two children in the ASD group had IGT. Moreover, four children in the ASD group showed a phenomenon of delayed insulin secretion.

Methods

Participants

Children with ASD were recruited from Snail Baby family fraternity. All ASD children participating in this study were diagnosed with Autism Diagnostic Observation Schedule (ADOS). The ADOS diagnostic procedure was performed by a qualified evaluator. Individual patients fulfilling the ADOS diagnostic procedure were included. The ADOS is designed to rate behaviors associated with ASD using a semi-structured, and observation-based approach. Each item assessing social communication or repetitive and stereotyped behaviors has a detailed operational definition describing the item theme [30]. Children who had neurological syndromes or focal neurological signs, fragile X syndrome, tuberous sclerosis, Down syndrome and other genetic diseases were excluded.

Control children were recruited from two kindergartens. Children who had psychiatric conditions (such as depressive disorder, attention deficit hyperactivity disorder, schizophrenia and bipolar disorder) were excluded according to their medical examinations for enrollment and parent interview. All participants had no family genetic history of diabetes and did not take medications (such as catecholamine, prazosin and sulfa drugs) in the past month.

At last, 17 children with ASD and 17 typically developing children were enrolled in Beijing between March 2016 and June 2017. The ages of autistic children and normal children were 4 to 8 years.

OGTT, insulin and C peptide release tests

To assess whether autism children emerge alterations in peripheral glucose metabolism, insulin sensitivity and beta cell function, OGTT, insulin and C peptide release tests were performed simultaneously. All participants were fasted for 10-12 hours prior to baseline glucose and then insulin and C peptide measurement were performed

with subsequent oral glucose (1.75 g/kg body weight, the total amount not exceeding 75g) in the morning. Venous blood samples were collected before (t=0), 30, 60 and 120 minutes after glucose administration in a grade A tertiary hospital in Beijing. The blood glucose concentration values were plotted against time to form a curve demonstrating the variations in blood glucose levels with time, defined as an integrated area of the curve for blood glucose. The same has been performed for the insulin and C-peptide. All blood samples at the four time points were collected from the children with ASD while only nine typically developing children completed all tests, with the remaining eight children had baseline data.

Homeostatic model assessment of insulin resistance (HOMA-IR) was calculated using homeostasis model assessment methods, defined as fasting insulin (IU/ml) *fasting glucose (mmol/L)/22.5. Homeostatic model assessment of insulin sensitivity (HOMA-IS) was defined as 22.5/ fasting insulin (IU/ml) *fasting glucose (mmol/L) [31].

Diagnostic criteria for IGT

According to the criteria of the American Diabetes Association, IGT is defined as: two-hour glucose levels of 140 to 199 mg per dl (7.8 to 11.0 mmol/l) on the OGTT [32]. A patient is said to be under the condition of IGT when he has an intermediately raised glucose level after two hours, but less than the level that would qualify for T2DM. The

fasting glucose may be either normal or mildly elevated.

Statistical analysis

Data were expressed as means ± SEM in this study. Statistical significance was calculated using two-sided independent sample t-tests or Pearson's two-sided chi squared test. The blood glucose, insulin and C peptide concentration were compared using a two-way ANOVA with repeated measures (2-way RM ANOVA) with sample time as the within-subjects variable (Graph Pad Prism version 6). Bonferroni post-hoc t-tests were performed to estimate the significance at diverse time-points of the glucose tolerance, insulin secretion and C peptide secretion curves. For all analyses, P values were two-tailed and statistical significance was set at P<0 .05.

Results

Demographic data and metabolic characteristics

In our study, there were no statistical differences in age, height, weight and BMI between ASD and control groups. However, maternal age of the ASD group was significantly higher than that of the control group.

There were no significant differences in fasting insulin, fasting C peptide, insulin resistance index and insulin sensitivity index. Table 1 showed detailed information about demographic data and metabolic characteristics.

	ASD group (n=17)	Control group (n=17)
	Mean (SEM)	Mean (SEM)
Sex	3 girls, 14 boys	3 girls, 14 boys
Age	5.1 (0.20)	5.3 (0.19) [#]
Height	110.9 (1.86)	109.3 (2.25)
Weight	19.7 (0.55)	19.3 (0.81) [#]
BMI	16.10 (0.40)	16.1 (0.28) [#]
Maternal age	30.4 (0.99)	26.4 (0.43) ^{**}
Fasting glucose (mmol/L)	4.5 (0.16)	5.0 (0.06) ^{**}

Fasting insulin (mU/L)	4.3 (0.75)	5.0 (0.58) [#]
Fasting C peptide (pmol/L)	193.52 (4.00)	244.8 (19.80) [#]
HOMA-IR ^a	0.9 (0.18)	1.1 (0.14) [#]
HOMA-IS ^b	1.8 (0.31)	1.2 (0.23) [#]
ADOS-TS ^c	22.2 (0.66)	-
ADOS-SCI ^d	16.3 (0.35)	-
^a Homeostatic model assessment of insulin resistance, ^b Homeostatic model assessment of insulin sensitivity, ^c Autism Diagnostic Observation Schedule-Total Score, ^d Autism Diagnostic Observation Schedule- Social communication and interaction, [#] Not significant, * <i>P</i> <0.05, ** <i>P</i> <0.01		

Table 1 : Group comparisons including sex, height, weight, BMI (Body Mass Index), maternal age and metabolic characteristics (continuous variables were represented by mean and Standard Error of Mean (SEM)).

Reduced fasting blood glucose in ASD children

The level of fasting blood glucose in the ASD group was significantly lower than that of the control group (*P*=0.0055). It might be related to the cognitive dysfunction in children with ASD. Besides, there were three children in the ASD group whose fasting blood glucose was below 4

mmol/L (Figure 1), suggesting insufficient energy supply of these children.

To determine whether the ASD children have altered peripheral glucose regulation, OGTT were performed on ASD and control groups. We found that two children with ASD had IGT according to 2-hour postprandial blood glucose level (8.4 and 8.7 mmol/L). IGT is a pre-diabetic state of hyperglycemia that is associated with insulin resistance. It is a strong predictor of T2DM. This result confirmed our initial speculation that the ASD children might exhibit a pre-diabetic phenotype during early childhood and were more likely to develop into T2DM in their adulthood. We found no IGT in the control group. However, chi-square analysis showed that the difference was statistically insignificant between ASD and control groups. The hallmark heterogeneity of ASD may be one of the reasons that we did not find statistical differences between the two groups. Nevertheless, our discovery may reflect the impaired glucose metabolism in a subgroup of ASD.

Besides, the results of our study revealed that there was no significant difference of glucose tolerance between two groups (Figure 2). Two-way ANOVA with repeated measures (2-way RM ANOVA) - group: *P*=0.6543, over time: *P*<0.0001, interaction: *P*=0.6499; AUC values (mean ± SEM) were: ASD 738.1 ± 35.54; Control 702.0 ± 27.52; *P*=0.5028). DM is a chronic disease and many

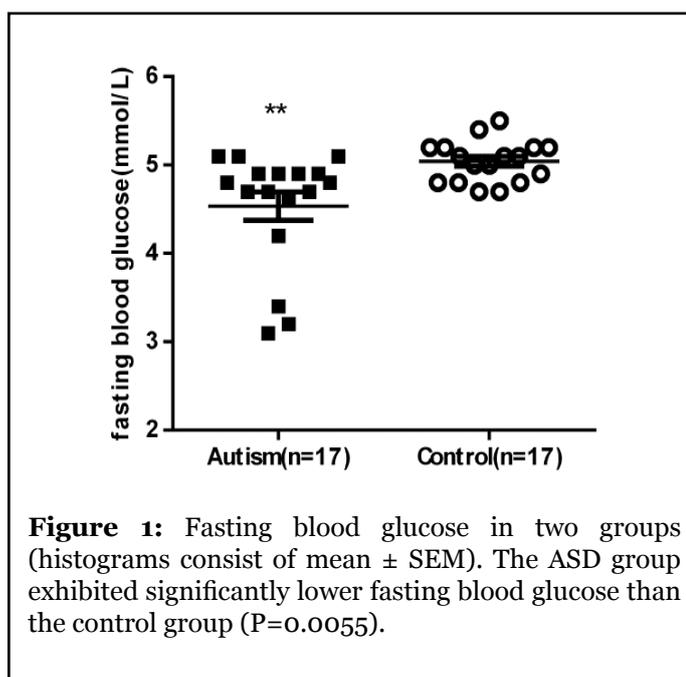
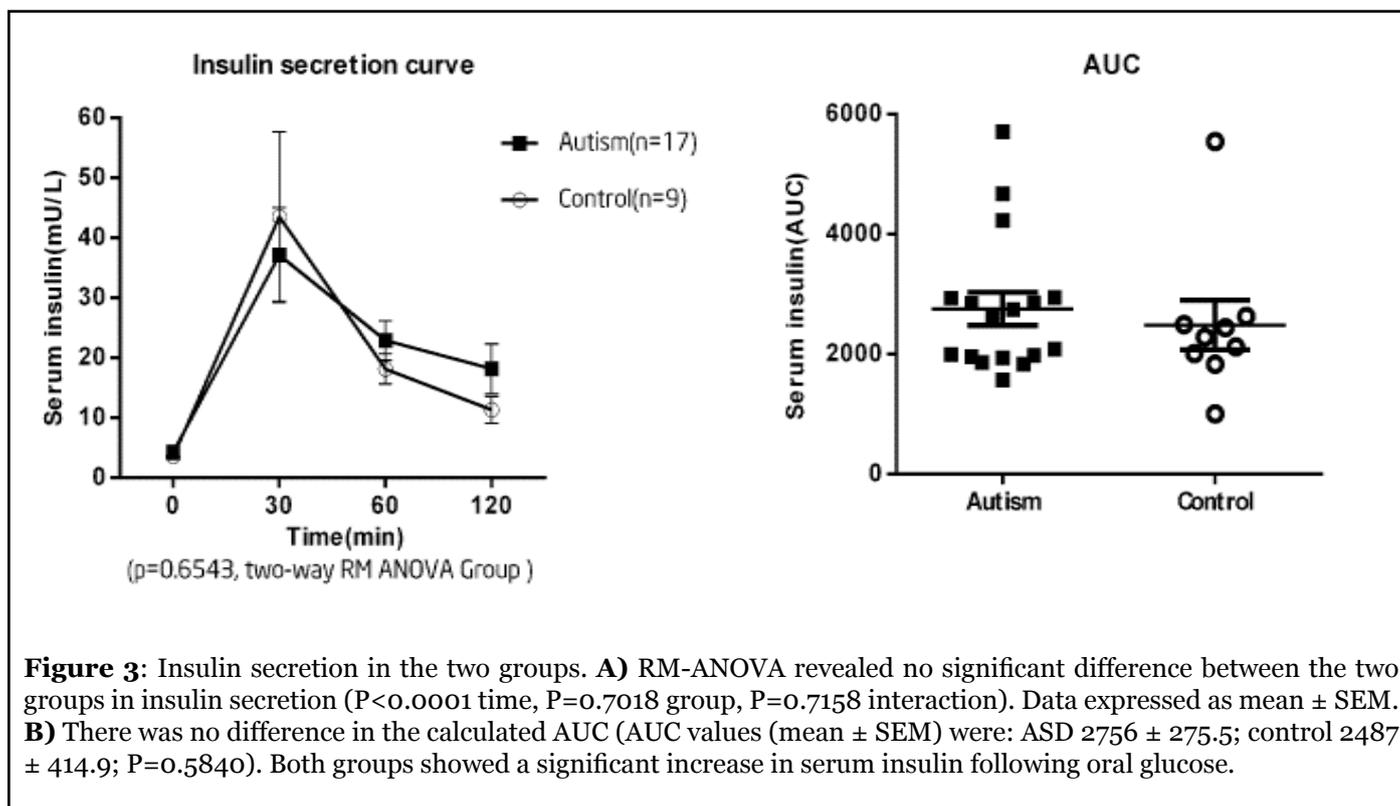
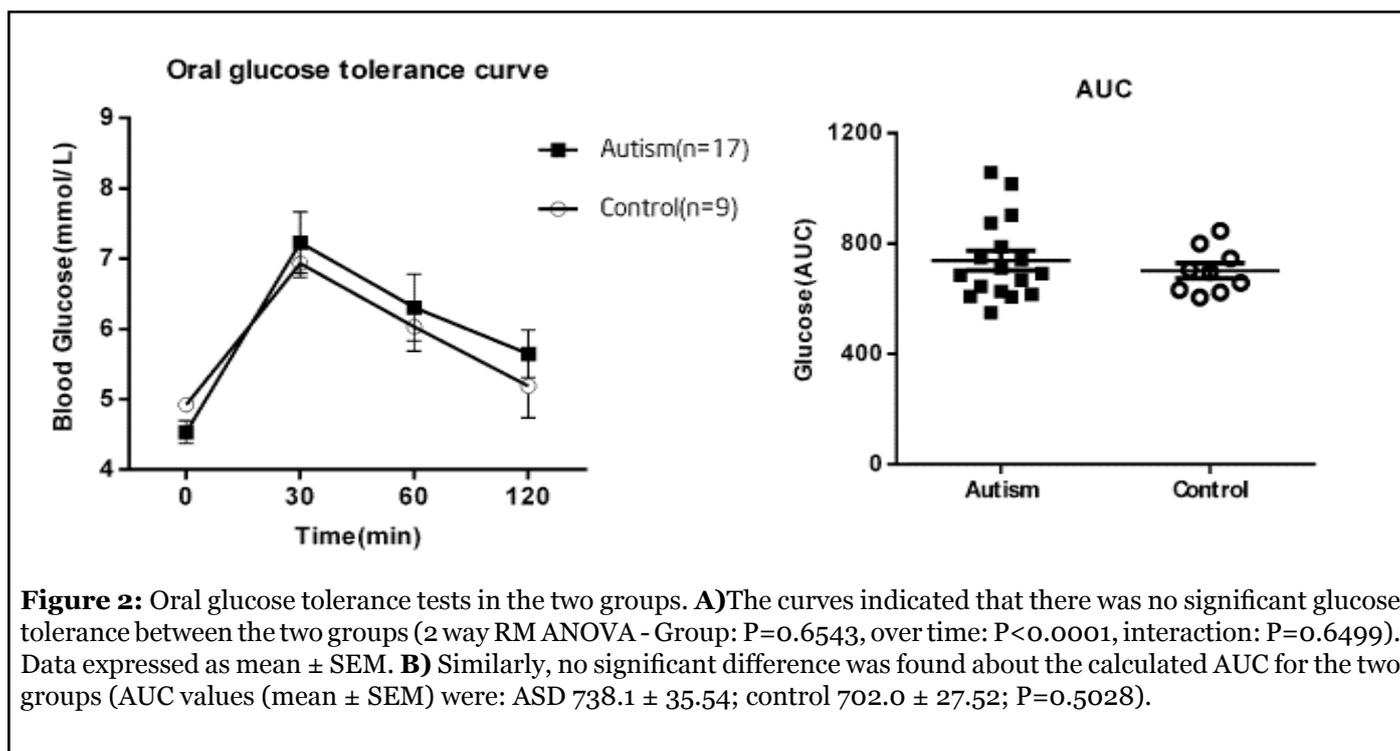


Figure 1: Fasting blood glucose in two groups (histograms consist of mean ± SEM). The ASD group exhibited significantly lower fasting blood glucose than the control group (*P*=0.0055).

factors involve in the development of it. To further unravel the difference of glucose tolerance between ASD and control groups, a larger sample size is needed.

To evaluate whether insulin secretion in the ASD group was changed, blood plasma samples were

collected during the OGTT at time 0, 30, 60 and 120 minutes to measure the insulin level. RM ANOVA was used to test the difference of insulin secretion between ASD and control groups. As a result, we found that oral glucose could cause increased plasma insulin in both ASD and control groups (Figure 3). RM ANOVA $P < 0.0001$ time, $P = 0.7018$ group, $P = 0.7158$



interaction; AUC values (mean ± SEM) were: ASD 2756 ± 275.5; control 2487 ± 414.9; P=0.5840). To compare the difference in insulin secretion, we assessed the percent change in insulin levels from time 0 to 30 minutes (Δ -insulin). The unpaired t-test analysis testified no statistical difference between groups in their capacity to release insulin (P=0.919), suggesting unchanged plasma insulin between children with ASD and control children. However, we found that four children in the ASD group showed a delayed insulin secretion (Figure 4). Their peak insulin secretion emerged at time point 120 minutes, suggesting that these children had defective early beta-cell function and a higher risk of developing diabetes. In addition, two children showed a delayed insulin secretion and lower fasting blood glucose. Thus, we speculated that the delayed insulin secretion might be linked to the lower fasting blood glucose.

Altered peak C-peptide level in ASD children

C-peptide is not destroyed by the liver, and the half-life period is much longer than that of the insulin. It can be an assessment for insulin secretion. In order to further explore the potential mechanism of IGT, we measured C-peptide levels before (t=0), 30, 60 and 120 minutes after glucose administration. The result of RM-ANOVA analysis pointed out that the plasma C-peptide was increased in both ASD and control groups after oral glucose (Figure 5). RM ANOVA

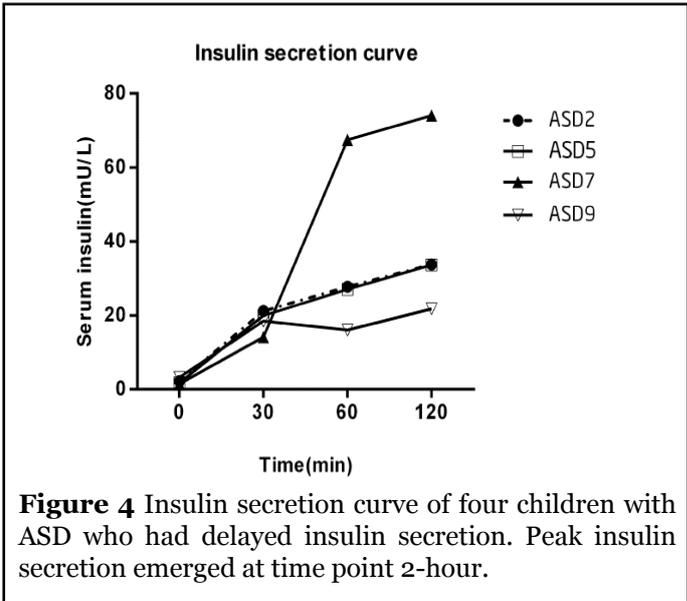


Figure 4 Insulin secretion curve of four children with ASD who had delayed insulin secretion. Peak insulin secretion emerged at time point 2-hour.

P<0.0001 time, P=0.2087 group, P=0.1154 interaction. AUC values (mean ± SEM) were: ASD 100258 ± 6457; Control 115862 ± 14197; P=0.2602). Besides, we found that the C-peptide level reached the peak at 30 minutes in both ASD and control groups, but the level in the ASD group was significantly lower than that in the control group (peak C-peptide level (mean ± SEM): ASD group, 1062.44 ± 139.97, control group: 1515.71 ± 231.66, P=0.0320). The reduced C-peptide level at 30 minutes indicated that there might be pancreatic β -cell dysfunction in children with ASD.

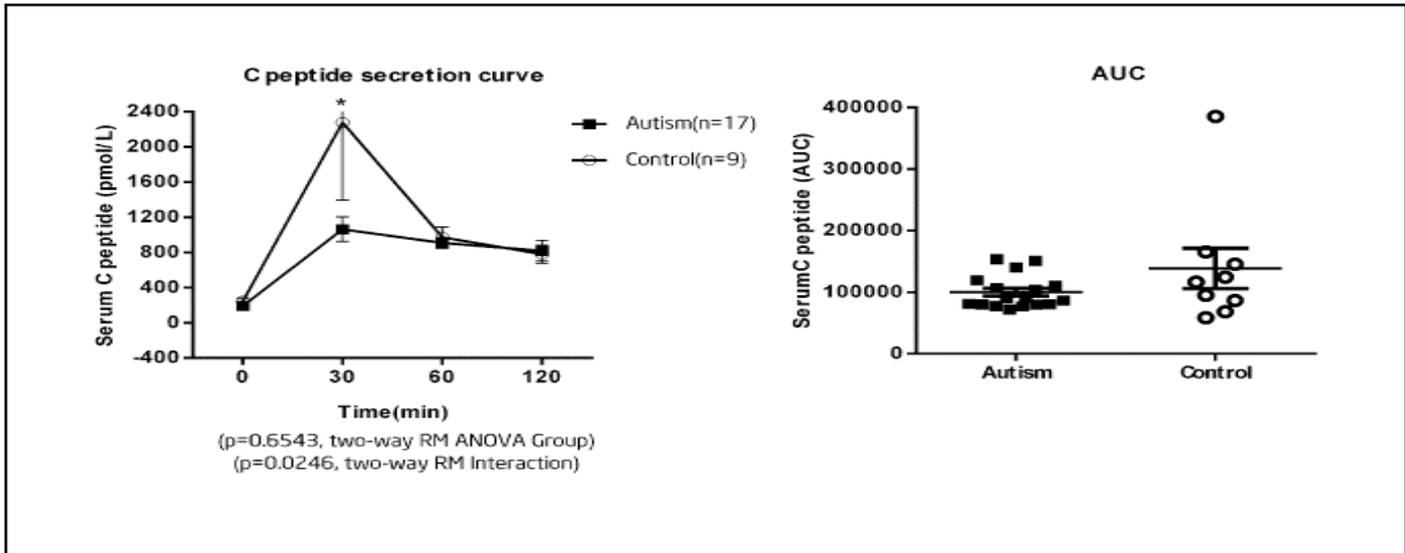


Figure 5: Comparison of C-peptide between the two groups. **A)** While a RM-ANOVA revealed no significant difference between the two groups of plasma C peptide secretion (P=0.2087), there was a significant difference in the interaction between time and group (RM ANOVA P<0.05). Indeed, plasma C peptide appeared to slightly increase, with a lower increase observed in the ASD group. Data expressed as mean ± SEM. **B)** There was no difference in the calculated AUC (AUC values (mean ± SEM) were: ASD group, 1062.44 ± 139.97, control group: 1515.71 ± 231.66, P=0.0320). Significance: *P<0.05.

Discussion

In our study, we found that fasting blood glucose concentration in the ASD group was lower than that of the control group. There were three children in the ASD group whose fasting blood glucose level was below 4 mmol/L. A previous study discovered that people with ASD had lower fasting blood glucose levels compared to a community-based population [14]. Low plasma glucose level was associated with poorer scores in attention tasks and memory [33,34]. It is well known that there is an equilibrium between plasma and brain glucose. Individuals with initially high levels of blood glucose may have higher levels of brain glucose [35]. A high level of baseline blood glucose was associated with better memory [36,37]. Participants who had a higher level of blood glucose solved the critical problems significantly faster than those with a lower blood glucose level [38]. The brain can't get enough glucose if the fasting blood glucose is too low. Clinical observation showed that ASD patients developed symptoms of working memory deficits, which severely disturb children's ability to learn [39]. Speculatively, lower fasting blood glucose may show altered glucose regulation in the ASD group and is related to the cerebral development. The abnormalities in regional cerebral glucose metabolism also appeared in patients with ASD [40].

We also observed that two children out of seventeen in the ASD group had IGT. We found no IGT in the control group. Besides, mean blood glucose level at 120 minutes in the ASD group was higher than that of the control group. IGT presents as a risk factor for the progress of DM and cardiovascular pathology [41]. It is a strong predictor of T2DM and the risk of T2DM significantly increased proportionally to two-hour glucose level on the OGTT [42]. The pre-diabetes found in ASD group might explain the higher incidence of T2DM in their later life. ASD had relations with low-grade inflammation state, marked by the increase of pro-inflammatory biomarkers [43]. The inflammation state can disturb insulin function and cause IGT [25]. However, chi-square analysis showed that the difference was statistically insignificant between ASD and control groups. Hence, big sample research is required to verify whether children with ASD show IGT.

Lower fasting blood glucose and IGT in children with ASD may be due to the alteration of pancreatic islet function. Four children in the ASD group showed a delayed insulin secretion; their peak insulin secretion emerged at time point 120 minutes. Normal glucose tolerance individuals with peak insulin secretion at 120 minutes had defective early beta-cell function and a higher risk of developing diabetes [44]. We also

demonstrated that the C-peptide level at 30 minutes in the ASD group was significantly lower than that of the control group ($P=0.0320$). The decreased C-peptide response in the ASD group might be a direct indicator of defective insulin secretion. Worsening glucose tolerance was associated with lower C-peptide [45]. Besides, there was a slight, although non-significant trend for decreased insulin level at 30 minutes in the ASD group. C-peptide has anti-inflammatory effects as well as an aid to repair smooth muscle cells [46,47]. While the relationship between DM and ASD is unclear, several factors probably contribute including: 1) similar common gene polymorphism, such as glyoxalase I (GLO1) C419A polymorphism [48], 2) related risk factors, such as dyslipidemia and obesity [49,50] and 3) altered microbial metabolites [51].

There are several additional unanswered questions that suggest future research. As the kinetics of immunoreactive insulin (IRI) and C-peptide differs considerably in human body, it is attracting to compare these two parameters under many pathophysiological conditions. Nevertheless, the temporal profile of C-peptide is always sluggish compared with that of insulin. We could not explain the discrepancy between the reduced C-peptide level and unchanged insulin level in children with ASD. Hence, more research is needed to study the pancreatic β -cell function of children with ASD.

Conclusion

We present the first trial of OGTT, insulin and C peptide release tests in children with ASD and control group. The results indicated that fasting blood glucose and C peptide level at 30 minutes significantly reduced in children with ASD. Two children in the ASD group had IGT according to 2-hour postprandial blood glucose level and four children in the ASD group emerged a phenomenon of delayed insulin secretion. All these results indicated that the pre-diabetic state may be a biological subtype of children with ASD. However, the causal relationship between ASD and DM is confusing. To further unravel the relationship between DM and ASD, research with larger sample is needed.

Conflicts of Interest

The authors declare that they have no competing interests.

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