

Hypertrophic Cardiomyopathy and the Troponins: The Enigma Remains

Rameen Shakur*

Janson Precision and Regenerative Medicine at MIT, The Koch Institute for Integrative cancer research, Massachusetts Institute of Technology, 500, Main street, Cambridge, MA, USA

*Correspondence should be addressed to Rameen Shakur; rshakur@mit.edu

Received date: January 11, 2021, **Accepted date:** March 10, 2021

Copyright: © 2020 Shakur R. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Hypertrophic cardiomyopathy (HCM) is a heart muscle disorder and is the most common form of Mendelian-inherited heart disease, affecting approximately 0.2% of the global population [1,2]. In adults the disease is often inherited as an autosomal dominant trait caused by mutations, mainly in one of the 23 cardiac sarcomere protein genes [3-5]. HCM is defined as left ventricular wall thickness or mass, in the absence of abnormal loading conditions [3]. Whilst, histologically it is characterised by myocardial disarray, fibrosis and small vessel disease [4]. HCM was the first of the cardiomyopathies to be attributed to a genetic aetiology, with 50% of HCM cases attributable to a specific disease-causing gene [4,6]. Validation studies through co-segregation and linkage analysis has identified only half of the 23 sarcomeric or sarcomeric associated proteins, but the majority of cases (almost three quarters) arise from 2 genes *MYH7* and *MYBPC3* [7]. A number of HCM causing mutations are unique to families known as 'private mutations'. The remainder of cases are found in the Thin filament complexes such as the Troponin T, I and Tropomyosin and rarely in the non sarcomeric proteins or metabolic genes such as PLN (Cardiac Phospholamban) *CAV3* (Caveolin-3) and *PRKAG2* (5'-AMP-activated protein kinase γ_2). Unfortunately, given the substantial allelic heterogeneity within each disease-causing gene and the large number of distinct mutations (>900) it remains a complex genetic conundrum [8,9]. This is further compounded by the variable disease penetrance, age of onset, and outcome. Hence, this would support the presence of other disease modifiers must exist either common or intermediate genetic variants across the entire genome, or interactions *via* epigenetic signalling. Molecularly a number of possible molecular pathogenesis for HCM have been postulated, amongst which impaired myofibrillar contractile function, perturbations in calcium sensitivity,

increased myocardial fibrosis are the main factors [10-13].

Impaired Myofibrillar Contractile Function

This was initially suggested to be the most-important mechanism, resulting in a 'compensatory' hypertrophy and diastolic dysfunction [14]. However, this generic mechanism of altered contractility caused by various sarcomeric gene mutations is not consistent across many of the common mutations. For example, mutations in *MYH7* have shown to produce either a reduced, or even enhanced, cardiac activity [15,16]. Meanwhile, other mutations in genes encoding thin-filament regulatory proteins, such as the troponins and α -tropomyosin, seem to frequently increase the calcium sensitivity of contractile proteins and so produce an augmented force of contraction within the confines of minimal calcium concentrations [17].

Impaired Calcium Sensitivity

Cardiac contraction is synonymous with calcium flux changes and the ongoing calcium cycling. Therefore, impaired calcium cycling be it from altered expression, phosphorylation or both from proteins such as sarcoplasmic/endoplasmic reticulum calcium ATPase 2 (SERCA) has been implicated in both systolic and diastolic heart failure [18]. This same premise has been shown in HCM [19]. Furthermore, experimental data from transgenic mice expressing the mutant form of human cardiac Troponin T with an associated HCM mutation by Knollmann et al. suggest that alterations in calcium cycling and homeostasis might also contribute to ventricular arrhythmias in absence of hypertrophy. Indeed, the cardiomyocytes from these mutant mice showed prolonged calcium transients compared to wild-type control mice,

which went on to trigger delayed after-depolarizations or spontaneous calcium oscillations [20]. This study provided the first *in vivo* evidence in the Troponin mutations that arrhythmogenesis was independent of structural changes in the myocardium; whilst elegantly showing a means to rescue this through Blebbistatin, which decreases calcium sensitivity. However, in contrast to this there has also been reports of reduced calcium sensitivity within the Troponin mutations but not specific to the *I79N* mutation [21].

Myocardial Fibrosis and HCM

The extent of scar and fibrosis has been shown to be correlated with reduced cardiac output and an increased propensity for heart failure and arrhythmogenesis in a HCM model with animals [22]. This is in support of a number of clinical *in vivo* studies using cardiac MRI and the application of late gadolinium enhancement as a surrogate for scar formation, which have for a while purported the relationship between fibrosis and poor clinical outcomes in HCM [23-25].

Yet, the molecular mechanism and trigger for this inherent process remains still an enigma. Some have identified the role of fibrosis to be attributed to apoptosis of myocytes, with the subsequent replacement by an expansion of the interstitial matrix individually leading onto cardiomyocyte hypertrophy, which may or may not be mutually exclusive [26]. In addition, further reports have shown fibrosis to be a mixture of pro-fibrotic molecules including collagens, elastin, with the main signalling pathway being through transforming growth factor β (TGF- β) [22]. This was confirmed when fibrosis was reduced following the administration of TGF- β -neutralizing antibody and Losartan, an angiotensin-II-receptor antagonist, given the role played by angiotensin in promoting TGF- β expression. Furthermore, the cardiac MRI data on fibrosis also highlighted areas of increased wall thickness, which implicated a relationship between cell turnover and growth of cardiomyocytes and fibrosis [27]. This was coupled with the imbalanced collagen turnover which was a result of the mismatch between extracellular-matrix synthesis and degradation hence leading onto the early histopathological remodelling that define HCM [28,29].

Sudden Death and HCM

The clinical phenotype for HCM is variable ranging from lifelong asymptomatic forms, dyspnea on exertion to early Sudden Cardiac Death (SCD). It is the most common cause of sudden cardiac death in individuals younger than 35 years of age and frequently those who are asymptomatic; importantly there appears to be an overall cumulating mortality of 1-2% per year for such patients [5,7]. The defining paradigm in SCD is the generation of dangerous if not fatal arrhythmias, the trigger for which is unclear,

but likely multifactorial including the development of non atherosclerotic induced hypoxic stress within the heart. The primary causative arrhythmia being either sustained Ventricular Tachycardia (VT) or Ventricular Fibrillation (VF), which is often initiated by premature ventricular complexes on a background of normal sinus rhythm [12]. Although there are many putative mechanisms for the induction of ventricular arrhythmias ranging from fibrosis to inefficient energy utilisation; a popular hypothesis for the induction of such arrhythmias has been increased alterations in calcium cycling homeostasis at a cardiomyocyte level and increased calcium sensitivity at the myofilament causing the generation of VT [12,13,27].

These effects were seen in the absence of hypertrophy, implying that arrhythmogenesis was not only due to macroscopic physical changes of the heart. Yet, the underlying molecular mechanisms as to how calcium induces ventricular arrhythmias and the signalling pathways remain unclear. Furthermore, the majority of such studies have taken place in the mice, which has distinct electrophysiological differences to humans; namely the high basal heart rate (300bpm), the very negative action potential (AP) plateau phase and the shorter duration of the AP (Figure 1).

Genetic Testing in HCM

Meanwhile, the prospect of genetic testing for risk prediction remains a rather mute topic. The initial analyses within large families of affected patients in single center's attempted to implicate the possibility of some mutations being associated with a more "malignant" course with a higher risk of sudden cardiac death and progressive heart failure [30]. However, in other large cohort and family studies, this was not always recapitulated [6]. Nevertheless, besides such conflicting data, some paradigms have been confirmed, namely involving the variable penetrance and clinical phenotype of particular mutations. One such gene is *TNNT2*, which is known to cause HCM but with little or even no hypertrophy. However, Individuals with these mutations seem to still possess a high risk of malignant ventricular arrhythmias and sudden cardiac death [8-10]. Given this intriguing disparity we decided to further investigate the Troponin and thin filament complexes and their interactions with each other in the context of clinical outcomes. This is particularly challenging given that sometimes it has been reported that certain mutations related to a HCM phenotype could also cause DCM and restrictive cardiomyopathies. This underlies the longstanding complexity between genotype-phenotype correlations. Yet, a full holistic and unbiased representation of the data, especially one which is able to integrate structural, genomic and clinical phenotypic data is lacking. In addition, the dynamic nature of cardiac contraction namely its effect with calcium and this interplay

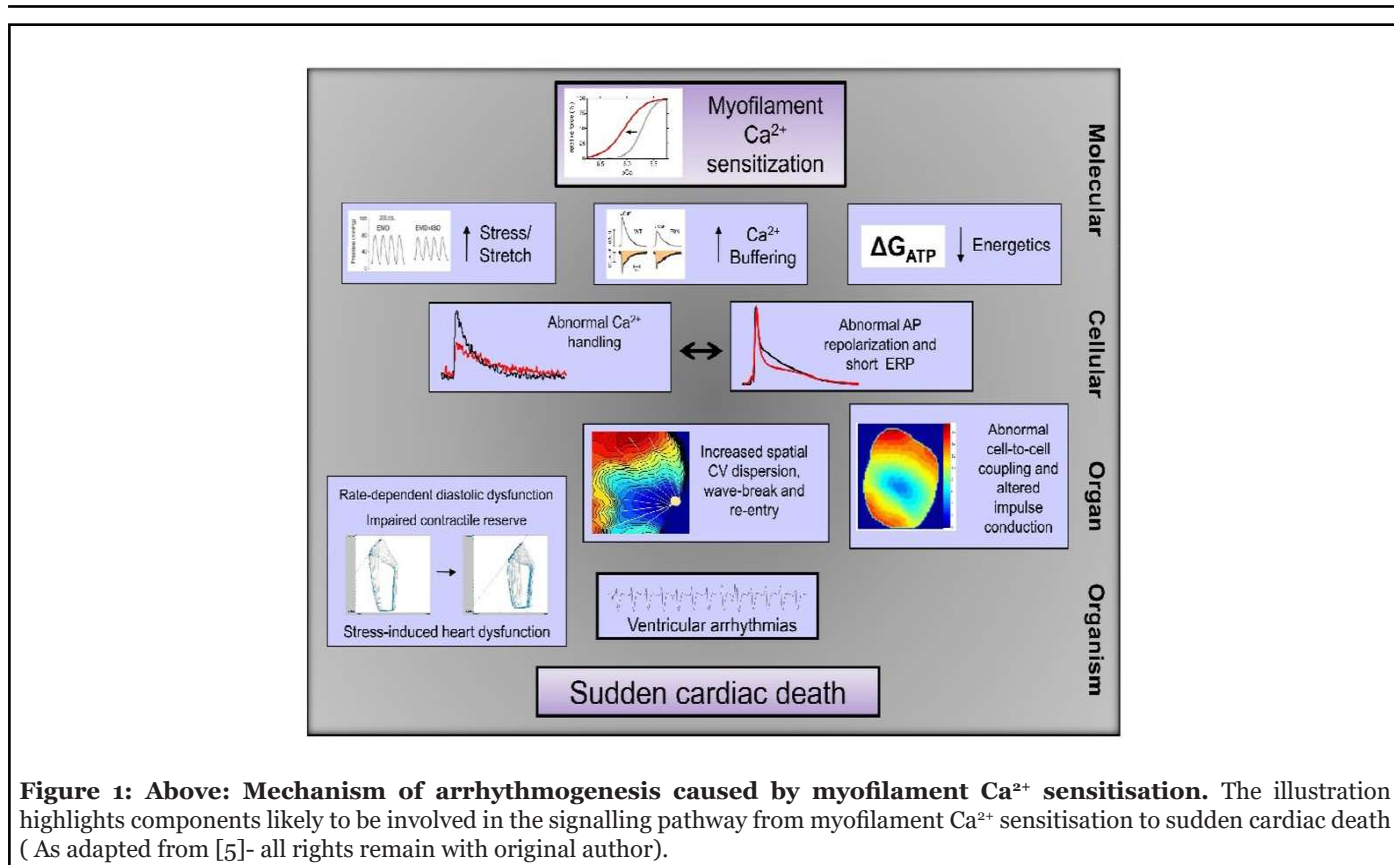


Figure 1: Above: Mechanism of arrhythmogenesis caused by myofilament Ca²⁺ sensitisation. The illustration highlights components likely to be involved in the signalling pathway from myofilament Ca²⁺ sensitisation to sudden cardiac death (As adapted from [5]- all rights remain with original author).

within such a model is also of importance to provide a more physiologically relevant model. The development of such a model will not only enhance our understanding of the genomic landscape in regards to disease ‘hot spots’ it may also provide a biological context prior to further experimental studies on mutations and their underlying mechanisms.

The Troponins

Since their discovery in 1963 the Troponin (Tn) proteins have been identified as part of a thin filament regulatory unit of the sarcomere. Troponin mutations are clinically relevant given they are thought to contribute approximately 15% of all the known sarcomeric protein cardiomyopathies. Unlike other such mutations it has been noted that the Troponin mutations they are however, show a more defined clinical course Troponin is a complex of three subunits. Troponin-I (Tn-I) inhibits Actomyosin ATPase; troponin-C (Tn-C) binds Calcium and troponin-T (Tn-T) links the complex to Tropomyosin (Tm) and is believed to be responsible for movement of Tm on the thin filament, modulating binding of the myosin head to Actin. The subunits are arranged in a 1:1:1 stoichiometric ratio along the thin filament with one Tn:Tm complex bound to every seven actin monomers. Troponin functions to couple calcium concentration changes to azimuthal movement of Tropomyosin on the thin filament. The position of Tropomyosin on actin controls the access of cross-bridges

to the thin filaments and thus regulates the cross-bridge cycling that drives contraction. At low calcium levels, Tropomyosin is held by troponin at a location that sterically blocks myosin binding sites on Actin, thus producing relaxation; this is the blocked or B state of the thin filament [26]. Thin filaments are switched on when Calcium binds to Troponin, which moves Tropomyosin to the closed or C-state position, where the myosin binding sites are partly uncovered. Myosin binding to the thin filament also alters the position of Tropomyosin, and full activation of the thin filament requires binding of both calcium and myosin. However, due to a lack of crystal structure data for the proteins namely Troponin T it has not been possible to infer the functional consequence of mutants in a robust manner and how a particular mutation may confer binding changes in relation to other members of the troponin complex such as to Troponin T and Troponin I.

Although by no means the most common causal genetic culprit for HCM, the prospect of further work in the field of mechanisms induced through the troponin complex mutations remain active. It is therefore foreseeable that the ongoing research in this area will have direct implications for therapeutic targets, clinical application and moreover supplement our more detailed molecular pathological understanding of this clinically devastating disease and one for which within cardiovascular disease remains the most promising for translational outcomes.

References

1. Bhatia RS, Tu JV, Lee DS, Austin PC, Fang J, Haouzi A, et al. Outcome of heart failure with preserved ejection fraction in a population-based study. *New England Journal of Medicine.* 2006 Jul 20;355(3):260-9.
2. Ashrafian H, Watkins H. Reviews of translational medicine and genomics in cardiovascular disease: new disease taxonomy and therapeutic implications: Cardiomyopathies: Therapeutics based on molecular phenotype. *Journal of the American College of Cardiology.* 2007 Mar 27;49(12):1251-64.
3. Elliott P, Andersson B, Arbustini E, Bilinska Z, Cecchi F, Charron P, et al. Classification of the cardiomyopathies: a position statement from the European Society Of Cardiology Working Group on Myocardial and Pericardial Diseases. *European Heart Journal.* 2008 Jan 1;29(2):270-6.
4. Varnava AM, Elliott PM, Sharma S, McKenna WJ, Davies MJ. Hypertrophic cardiomyopathy: the interrelation of disarray, fibrosis, and small vessel disease. *Heart.* 2000 Nov 1;84(5):476-82.
5. Maron BJ, Shirani J, Poliac LC, Mathenge R, Roberts WC, Mueller FO. Sudden death in young competitive athletes: clinical, demographic, and pathological profiles. *JAMA.* 1996 Jul 17;276(3):199-204.
6. Tester DJ, Ackerman MJ. Genetic testing for potentially lethal, highly treatable inherited cardiomyopathies/channelopathies in clinical practice. *Circulation.* 2011 Mar 8;123(9):1021-37.
7. Elliott PM, Poloniecki J, Dickie S, Sharma S, Monserrat L, Varnava A, et al. Sudden death in hypertrophic cardiomyopathy: identification of high risk patients. *Journal of the American College of Cardiology.* 2000 Dec;36(7):2212-8.
8. Nicod P, Polikar R, Peterson KL. Hypertrophic cardiomyopathy and sudden death. *New England Journal of Medicine.* 1988 May 12;318(19):1255-7.
9. Marston SB. How do mutations in contractile proteins cause the primary familial cardiomyopathies?. *Journal of Cardiovascular Translational Research.* 2011 Jun;4(3):245-55.
10. Maron BJ, Olivotto I, Spirito P, Casey SA, Bellone P, Gohman TE, et al. Epidemiology of hypertrophic cardiomyopathy-related death: revisited in a large non-referral-based patient population. *Circulation.* 2000 Aug 22;102(8):858-64.
11. Maron BJ, Spirito P, Shen WK, Haas TS, Formisano F, Link MS, et al. Implantable cardioverter-defibrillators and prevention of sudden cardiac death in hypertrophic cardiomyopathy. *JAMA.* 2007 Jul 25;298(4):405-12.
12. Knollmann BC, Kirchhof P, Sirenko SG, Degen H, Greene AE, Schober T, et al. Familial hypertrophic cardiomyopathy-linked mutant troponin T causes stress-induced ventricular tachycardia and Ca²⁺-dependent action potential remodeling. *Circulation Research.* 2003 Mar 7;92(4):428-36.
13. Baudenbacher F, Schober T, Pinto JR, Sidorov VY, Hilliard F, Solaro RJ, et al. Myofilament Ca²⁺ sensitization causes susceptibility to cardiac arrhythmia in mice. *The Journal of Clinical Investigation.* 2008 Dec 1;118(12):3893-903.
14. Revera M, Van Der Merwe L, Heradien M, Goosen A, Corfield VA, Brink PA, et al. Troponin T and β -myosin mutations have distinct cardiac functional effects in hypertrophic cardiomyopathy patients without hypertrophy. *Cardiovascular Research.* 2008 Mar 1;77(4):687-94.
15. Sata M, Ikebe M. Functional analysis of the mutations in the human cardiac beta-myosin that are responsible for familial hypertrophic cardiomyopathy. Implication for the clinical outcome. *The Journal of Clinical Investigation.* 1996 Dec 15;98(12):2866-73.
16. Lowey S. Functional consequences of mutations in the myosin heavy chain at sites implicated in familial hypertrophic cardiomyopathy. *Trends in Cardiovascular Medicine.* 2002 Nov 1;12(8):348-54.
17. Michele DE, Albayya FP, Metzger JM. Direct, convergent hypersensitivity of calcium-activated force generation produced by hypertrophic cardiomyopathy mutant α -tropomyosins in adult cardiac myocytes. *Nature Medicine.* 1999 Dec;5(12):1413-7.
18. Wehrens XH, Marks AR. Novel therapeutic approaches for heart failure by normalizing calcium cycling. *Nature Reviews Drug Discovery.* 2004 Jul;3(7):565-74.
19. Bottinelli R, Coviello DA, Redwood CS, Pellegrino MA, Maron BJ, Spirito P, et al. A mutant tropomyosin that causes hypertrophic cardiomyopathy is expressed in vivo and associated with an increased calcium sensitivity. *Circulation Research.* 1998 Jan 23;82(1):106-15.
20. Knollmann BC, Kirchhof P, Sirenko SG, Degen H, Greene AE, Schober T, et al. Familial hypertrophic cardiomyopathy-linked mutant troponin T causes stress-induced ventricular tachycardia and Ca²⁺-dependent action potential remodeling. *Circulation Research.* 2003 Mar 7;92(4):428-36.
21. Sweeney HL, Feng HS, Yang Z, Watkins H. Functional analyses of troponin T mutations that cause hypertrophic cardiomyopathy: insights into disease pathogenesis and troponin function. *Proceedings of the National Academy of Sciences.* 1998 Nov 24;95(24):14406-10.
22. Teekakirikul P, Eminaga S, Toka O, Alcalai R, Wang

- L, Wakimoto H, et al. Cardiac fibrosis in mice with hypertrophic cardiomyopathy is mediated by non-myocyte proliferation and requires Tgf- β . *The Journal of Clinical Investigation.* 2010 Oct 1;120(10):3520-9.
23. Varnava AM, Elliott PM, Mahon N, Davies MJ, McKenna WJ. Relation between myocyte disarray and outcome in hypertrophic cardiomyopathy. *The American Journal of Cardiology.* 2001 Aug 1;88(3):275-9.
24. Moon JC, Reed E, Sheppard MN, Elkington AG, Ho S, Burke M, et al. The histologic basis of late gadolinium enhancement cardiovascular magnetic resonance in hypertrophic cardiomyopathy. *Journal of the American College of Cardiology.* 2004 Jun 16;43(12):2260-4.
25. O'Hanlon R, Grasso A, Roughton M, Moon JC, Clark S, Wage R, et al. Prognostic significance of myocardial fibrosis in hypertrophic cardiomyopathy. *Journal of the American College of Cardiology.* 2010 Sep 7;56(11):867-74.
26. Konno T, Chen D, Wang L, Wakimoto H, Teekakirikul P, Naylor M, et al. Heterogeneous myocyte enhancer factor-2 (Mef2) activation in myocytes predicts focal scarring in hypertrophic cardiomyopathy. *Proceedings of the National Academy of Sciences.* 2010 Oct 19;107(42):18097-102..
27. Shivakumar K, Dostal DE, Boheler K, Baker KM, Lakatta EG. Differential response of cardiac fibroblasts from young adult and senescent rats to ANG II. *American Journal of Physiology-Heart and Circulatory Physiology.* 2003 Apr 1;284(4):H1454-9.
28. Ho CY, López B, Coelho-Filho OR, Lakdawala NK, Cirino AL, Jarolim P, et al. Myocardial fibrosis as an early manifestation of hypertrophic cardiomyopathy. *New England Journal of Medicine.* 2010 Aug 5;363(6):552-63.
29. Lombardi R, Betocchi S, Losi MA, Tocchetti CG, Aversa M, Miranda M, et al. Myocardial collagen turnover in hypertrophic cardiomyopathy. *Circulation.* 2003 Sep 23;108(12):1455-60.
30. Van Driest SL, Ackerman MJ, Ommen SR, Shakur R, Will ML, Nishimura RA, et al. Prevalence and severity of "benign" mutations in the β -myosin heavy chain, cardiac troponin T, and α -tropomyosin genes in hypertrophic cardiomyopathy. *Circulation.* 2002 Dec 10;106(24):3085-90.